

REVIEW ARTICLE

Open Access



Oxidative Stress and Its Biomarkers in Cardiovascular Diseases

Julija Valaitienė¹ and Agnė Laučytė-Cibulskienė^{2,3*} 

Abstract

Background Cardiovascular diseases (CVDs) are the most common cause of death worldwide. CVDs share heterogeneous pathophysiologic mechanisms, one of which includes increased oxidative stress.

Main Body Surplus levels of reactive oxygen species induce damage to cellular macromolecules such as DNA, proteins, and lipids. Increased reactive oxygen species result in decreased nitric oxide availability, vasoconstriction, and the development of procoagulant and proinflammatory states in blood vessels.

Conclusion Improved knowledge of biomolecular processes triggered by oxidative stress has helped develop tools for assessing oxidative stress markers and applying them in clinical settings. Nevertheless, some research gaps should be filled, specifically by defining the most clinically relevant biomarkers for oxidative stress with high sensitivity and specificity for CVD.

Keywords Biomarkers, Cardiovascular diseases, Endothelial cell dysfunction, Inflammation, Oxidative stress, Reactive oxygen species

1 Background

Cardiovascular diseases (CVDs) remain a top global issue [1] despite numerous initiatives to reduce their prevalence and impact on human health. The most prevalent type of cardiovascular disease (CVD) is coronary heart disease (CHD) [2]. Research evidence indicates that oxidative stress is a significant factor in the development of CVDs [3]. Significant oxidative stress leads to dysfunction and inflammation in blood vessels, primarily affecting endothelial cells (EC). Other blood vessel cells, such as vascular smooth muscle cells (VSMCs) or adventitia cells, are also involved [4]. However, ECs play a critical role in cardiovascular imbalance, e.g., endothelial

dysfunction impairs vasoconstriction and vasodilatation, causes EC apoptosis, increases the adhesion of ECs to monocytes, and alters the angiogenic potential of ECs [5]. Consequently, atherosclerotic plaques and lesions form and thus lead to CVD.

Excessive production or accumulation of reactive oxygen species (ROS) contributes to oxidative stress. ROS include the oxygen free radicals superoxide, hydroxyl, and peroxy radicals and nonradicals (hydrogen peroxide and hypochlorous acid) [6]. Mitochondria are the primary drivers of intracellular oxidant production in most cell types [7], followed by sources such as nicotinamide adenine dinucleotide phosphate oxidases (NOXs), heme oxygenase 1, xanthine oxidase, and cyclooxygenases [8, 9]. Basal levels of ROS generation are essential for signal transduction pathways, protection against microorganisms, gene expression, and the promotion of cellular survival, apoptosis or death [10, 11]. The body has protective measures against ROS, including enzymatic compounds such as glutathione peroxidase, superoxide dismutase (SOD),

*Correspondence:

Agnė Laučytė-Cibulskienė
agne.laucyte-cibulskiene@med.lu.se

¹ Faculty of Medicine, Vilnius University, Vilnius, Lithuania

² Department of Nephrology, Skåne University Hospital, Ruth Lundskogs Gata 10, 205 02 Malmö, Sweden

³ Department of Clinical Sciences, Lund University, Malmö, Sweden



© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

and catalase, as well as nonenzymatic compounds such as nicotinamide, glutathione and tocopherol [12–15].

Identifying numerous biomarkers for oxidative stress has made it easier to measure the levels of ROS. However, the use of these biomarkers in clinical settings still requires validation because of the considerable variation in oxidative stress levels across different diseases [16]. A biomarker is any substance or process that can be measured in the body or its products and can predict the occurrence of a disease or its outcome [17]. Many markers for oxidative stress can be measured, but their clinical applicability is a concern since there is no consensus on which one is superior to others. For a biomarker to be clinically beneficial, it must meet at least one of the following criteria: (a) demonstrate specificity for a particular disease, (b) have prognostic value, and (c) be correlated with disease activity [16]. This review also highlights the use of currently employed drugs with cardioprotective effects, such as sodium–glucose transport protein 2 (SGLT2), mineralocorticoid receptor antagonists, and glucagon-like peptide-1 (GLP-1) agonists, and their possible effects on oxidative stress.

This study investigated the connection between oxidative stress and endothelial cell dysfunction, particularly the impact of reduced nitric oxide (NO) bioavailability and inflammation caused by ROS. Furthermore, the biomarkers utilized to assess oxidation-specific epitopes (endogenous damage-associated molecular patterns) in CVD as indicators of oxidative stress and currently used drugs with antioxidant effects are reviewed.

Key terms

CVD – refers to atherosclerotic cardiovascular disease due to plaque buildup in artery walls, such as coronary artery disease, including acute coronary syndrome (myocardial infarction) and chronic coronary disease, cerebrovascular disease (stroke, transient ischemic attack, carotid artery stenosis), peripheral artery disease, abdominal and thoracic aortic aneurysm, and intestinal ischemia

Atherosclerosis – condition when cholesterol, fat, blood cells and other substances in blood form plaque on artery wall, leading to artery narrowness

Necrotic core – early stage of atherosclerosis defined by macrophage apoptosis and diminished clearance of apoptotic cells

EC – endothelial cells – cells that are located in tunica intima of blood vessels

Plaque formation – atherosclerotic plaque formation involving low density lipoprotein (LDL) accumulation in tunica intima, oxidation of LDL, recruitment of monocytes-macrophages, uptake of oxidized LDL by macrophages and transformation of macrophages into foam cells

ROS – reactive oxygen species, including free radicals as superoxide, hydroxyl, and peroxy radicals, and nonradical as hydrogen peroxide and hypochlorous acid

2 The Role of the Endothelium

Oxidative stress is the primary mechanism that provokes endothelial dysfunction characterized by the procoagulant, proinflammatory, and proliferative phenotype of ECs, resulting in atherothrombosis and coexisting inflammation [18, 19]. Additionally, endothelial dysfunction can be directly caused by increased levels of fatty acids in the blood, which induce insulin resistance, activate the renin–angiotensin system, and maintain inflammation [18, 20].

Excess free radicals reduce the bioavailability of endothelium-secreted vasodilators such as nitric oxide (NO) and prostacyclin (PGI₂) [21, 22]. In particular, NO has anti-inflammatory, antithrombotic properties (inhibits platelet aggregation) and protects blood vessels from vasospasm (acts through guanylate cyclase located in the membrane of vascular smooth muscle cells) [23]. However, NO is inactivated during the reaction with superoxide anions, forming peroxynitrite ONOO[−]. Free radicals also uncouple NO synthase (eNOS), thus decreasing its efficacy and the concentration of its substrates and cofactors (L-arginine and tetrahydrobiopterin BH₄, respectively) and increasing the level of the endogenous eNOS inhibitor dimethylarginine (Fig. 1) [19, 24]. Moreover, free radicals monomerize eNOS dimers, significantly reducing the efficiency of NO synthesis. ONOO[−] nitrates cellular proteins, including those in the electron transport chain, leading to mitochondrial dysfunction and cell apoptosis [19]. The hallmark of mitochondrial dysfunction is disturbed mitochondrial Ca²⁺ ion homeostasis (ROS inhibit mitochondrial Na⁺/Ca²⁺ exchangers) and altered membrane potential [25]. The mitochondrion itself also produces ROS, which cause mitochondrial DNA (mtDNA) damage [26] and contribute to endothelial dysfunction [27]. Endogenous and exogenous ROS act through mitogen-activated protein kinases (MAPKs) and increase the concentration of proliferative molecules such as fibroblast growth factor, insulin-like growth factor, platelets, and epidermal growth factor and the expression of their receptors in the vascular smooth muscle layer, which causes smooth muscle cells to migrate toward the endothelium and proliferate [28, 29]. Additionally, a dysfunctional endothelium initiates and maintains coagulation conditions through secreted von Willebrand factor (vWF), which interacts with platelet GPIIb/IIIa receptors, tissue factors, and plasminogen activator inhibitors [19].

3 The Role of Inflammation and Other Cells

During oxidative stress, ROS activate nuclear factor κ B (NF- κ B), which turns on target genes responsible for producing adhesion molecules (P and E selectins,

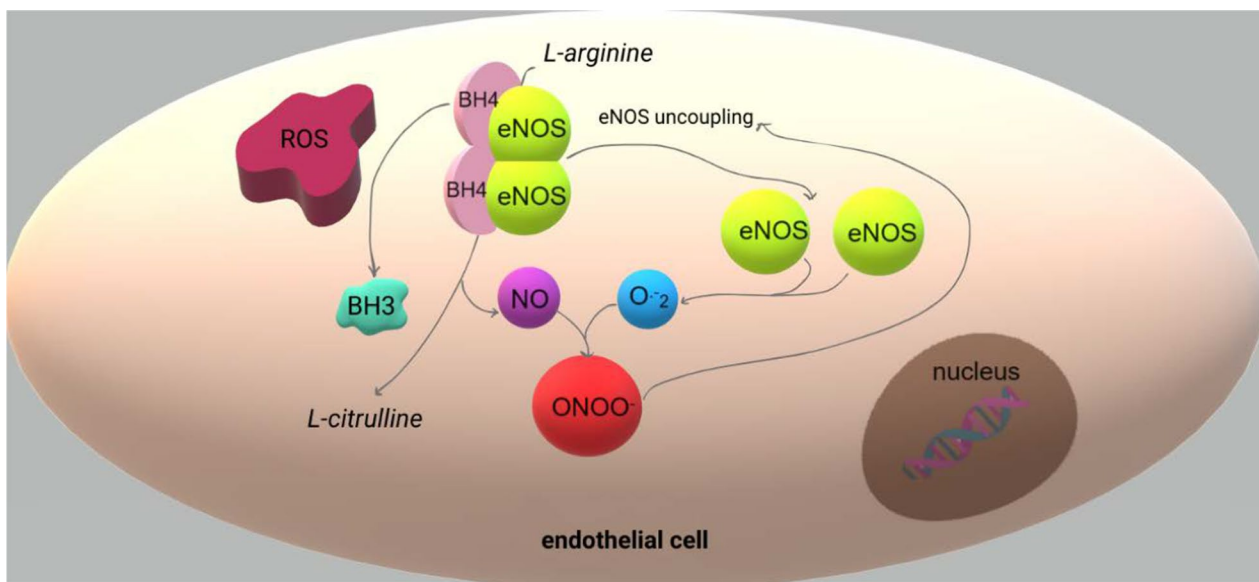


Fig. 1 Oxidative stress induced the uncoupling of eNOS in endothelial cells

ICAM-1, VCAM-1), IL-1, IL-6, TNF- α , and monocyte chemoattractant protein-1 (MCP-1). As a consequence, neutrophils and monocytes are activated and migrate to damaged blood vessel tissues (Fig. 2) [30, 31]. Dysfunctional endothelium also secretes extracellular vesicles with microRNA155 (miR155) and miR92a, which induce monocyte polarization toward a proinflammatory M1

phenotype [30]. Additionally, in response to IL-1 β , ox-LDL promotes the structural and functional transition of smooth muscle cells to cells with a macrophage phenotype [32, 33]. Activated neutrophils begin to secrete cathepsin G, which acts on platelets such as thrombin (through PAR1 receptors) and tissue factor, triggering the coagulation cascade [34].

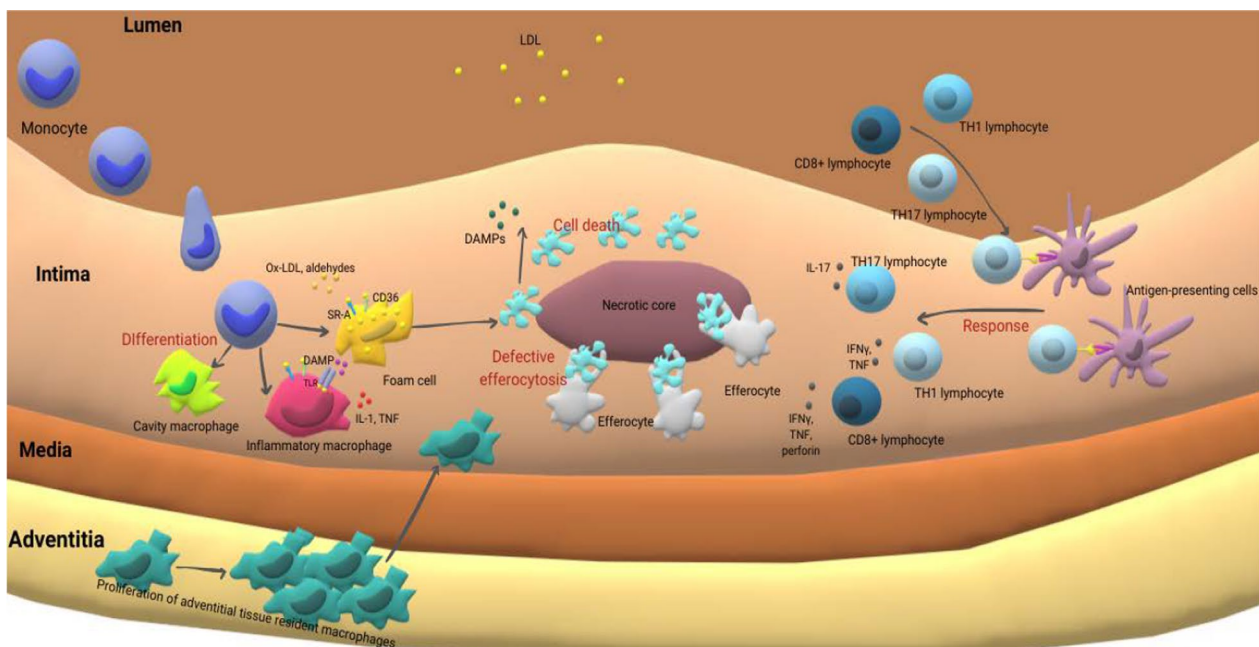


Fig. 2 Mechanisms of atherosclerotic plaque formation. The role of inflammation

Similarly, platelets can also activate neutrophils through their granule components (CXCL4 and HMGB1), stimulating the neutrophil receptors Mac1, PSGL-1, and CD40 [34, 35]. Neutrophils form and secrete neutrophil extracellular traps (NETs) after being stimulated by ox-LDL, cholesterol crystals, activated platelets, and IL-8 [36].

Consequently, NETs stimulate macrophages to produce IL-1 β cytokines, further contributing to the migration of neutrophils and T lymphocytes [37]. The latter secrete IL-17 and activate adaptive immunity [36]. When monocytes turn into macrophages, they can absorb lipoproteins via receptors such as CD36, SRA1, and SRA2. This process can cause the transformation of macrophages into foam cells. Furthermore, the expression of the pro-inflammatory endothelial adhesion molecules netrin-1 and semaphorin-3E restricts macrophage migration from atherosclerotic lesions and blocks the cytokine receptors CCL19 and CCL21, thus inhibiting macrophage chemotaxis [38, 39]. The subsequent retention of macrophage migration and ineffective efferocytosis lead to the formation of a necrotic lipid core with dead foam cells and efferocytes [39]. Mature dendritic cells, triggered by inflammatory cytokines and Toll-like receptor (TLR) agonists, begin to express CD11c+, CD11b+, and CD40 receptors and present antigens to T lymphocyte subtypes (CD8+ T lymphocytes through MHC class I molecules and to CD4+ through MHC class II molecules) [40–43]. Activated T lymphocytes (helper TH1, TH17 and cytotoxic CD8+ T cells) further increase inflammation (secretion of IFN γ , TNF, and IL-17) and stimulate vascular smooth muscle cell proliferation.

4 Oxidative Stress and Cardiovascular Disease

Oxidative stress is one of the most critical components in the pathogenesis of CVD, triggering thromboinflammation [44, 45]. It is caused by the excessive production of free radicals (ROS, nitrogen, and sulfur) or insufficiency of the antioxidant system [19, 46]. ROS include the hydrogen peroxide H₂O₂, superoxide anion O₂[−], and hydroxyl-OH [19]. They are byproducts of mitochondrial metabolism but can also be generated by the action of heme oxygenase 1, xanthine oxidase, and NADPH oxidases (NOXs) [47–49]. The mediators of oxidative stress in CVD include oxidized low-density lipoproteins (ox-LDL), angiotensin II, endothelin I, aldosterone, and glycosylated compounds. They bind to lectin-oxidized LDL receptor-1 (LOX-1) and activate NADPH oxidases (NOXs) [18, 29, 50–53]. Notably, increased blood pressure results in increased expression of LOX-1 and upregulated NOXs (mainly NOX1 and NOX4), leading to decompensated oxidative stress and the development of atherosclerosis in individuals with arterial hypertension [29, 54, 55].

Endothelial dysfunction may lead to the rupture of an atherosclerotic plaque and thus clot formation. The risk of rupture is significantly increased if one of the following conditions are present: a large necrotic core of the plaque, a thin fibrin cap (<65 μ m), pronounced inflammation, or vascular remodeling [56]. As a consequence, macrophages express fibrin cap-lysing enzymes (collagenases and gelatinases) and, together with T lymphocytes, secrete IFN γ , which inhibits collagen synthesis and induces VSMC apoptosis [57, 58].

VSMCs activate platelets through C-type lectin-like receptor 2 (CLEC-2), which has a very similar function to the collagen receptor GP VI (which also induces platelet activation) [59]. CLEC-2 is a newly identified protein on the surface of platelets [59]. CLEC-2 and podoplanin, endogenous ligands of CLEC-2, are both expressed in advanced atherosclerotic lesions. However, in early atherosclerotic lesions, only CLEC-2-binding sites are colocalized within VSMCs, whereas podoplanin expression is absent [60].

After atherosclerotic plaques rupture, large amounts of collagen, tissue factor, and vWF are released [61, 62]. vWF binds to the GP Ib–IX–V receptor complex of platelets and collagen to GP Ia/IIa and VI receptors, thus activating platelets and changing their shape and granule contents (ADP, serotonin, thromboxane A₂), leading to platelet adhesion [63, 64]. The release of tissue factor initiates the coagulation cascade via the extrinsic pathway and fibrin production, causing blood vessel thrombosis and, thus, CVD and possibly death.

5 Biomarkers of Oxidative Stress in Cardiovascular Diseases

There are numerous biomarkers for oxidative stress in CVD. Nevertheless, their clinical applicability is a concern, mostly because no consensus exists on which method is superior. Hence, only biomarkers specific for CVD are discussed below.

5.1 Protein Carbonyls and Advanced Glycation End Products

Protein carbonyls are created in several ways, namely, (a) through the oxidative breakdown of the protein skeleton; (b) through the coupling of aldehydic lipid peroxidation products to lysine, cysteine, and histidine remnants and the production of advanced lipoxidation end products; and (c) through the nonenzymatic glycation of reducing sugars [65–67]. Similarly, advanced glycation end products (AGEs) are formed via reactions between lysine and arginine residues and carbohydrates [16]. AGEs appear throughout normal metabolism but are more highly expressed in oxidative stress and hyperglycemic or hyperlipidemic states [65]. Consequently,

Table 1 Associations between oxidative stress biomarkers and cardiovascular diseases in human studies

Biomarker	Method	Change	Results	Pathology
Protein carbonyls [73, 74]	Reaction of the carbonyl group with 2,4-dinitrophenylhydrazine to form the 2,4-dinitrophenylhydrazone	↑	Cut-off for ACS 2.21 mmol/mg, AUC 0.84 (specificity 80%, sensitivity 83%)	Unstable angina, non-ST and ST elevation myocardial infarction
Advanced oxidation protein products [75]	Enzyme-linked-immunosorbent assay	↑	Abdominal aortic aneurysms ($r=0.4180$, $p\leq 0.05$), aortoliac occlusive disease ($r=0.616$, $p\leq 0.05$)	Abdominal aortic aneurysms, aortoliac occlusive disease
Oxidized low-density lipoproteins (oxidized phospholipids) [90, 91]	Enzyme-linked-immunosorbent assay	↑	Cardiovascular diseases (Effect Size total 0.75 (95% CI: 0.46, 1.03)	Cardiovascular diseases
Trans-4-hydroxy-2-nominal [77, 103]	Enzyme-linked immunosorbent assay, co-immunoprecipitation, immunoblot/Western blot	↑	Acute cardiovascular events (myocardial infarction, ischemic stroke), heart failure (HR: 2.23 (95% CI 1.44, 3.44); $p=0.0003$)	Acute cardiovascular events (myocardial infarction, ischemic stroke), heart failure
F ₂ -isoprostanes [113]	Enzyme-linked immunosorbent assay	↑	Coronary heart disease (OR: 95% CI 2.47 (1.44, 4.26), $p=0.001$), ischemic stroke (medians 0.041 vs. 0.0295, $p=0.012$; AUC=0.68 (0.55–0.8)	Coronary heart disease, ischemic stroke
Malondialdehyde [114]	Enzyme-linked immunosorbent assay	↑	Chronic heart failure (HR: 1.90 (95% CI 0.99, 3.65), $p=0.05$), history of myocardial infarction (HR: 1.64 (95% CI 1.00, 2.68), $p=0.05$), multivessel disease (HR: 1.78 (95% CI 1.09, 2.91), $p=0.02$)	Chronic heart failure history of myocardial infarction multivessel disease

AGEs interact with macrophages and endothelial cells, induce inflammation in an NF- κ B-dependent fashion, and trigger the expression of adhesion molecules and procoagulant tissue factors on the endothelium [68, 69]. AGEs are associated with arteriosclerosis, diabetes mellitus, and other conditions, such as obesity and neurodegenerative diseases [70]. Glycoxidation is another process that results in the generation of AGEs [16, 71, 72]. The essential clinically valuable feature of protein carbonyls is their stability in blood for up to 18 h [16]. Protein carbonyls are identified primarily after the derivatization of 2,4-dinitrophenylhydrazine (DNP) [65, 73]. With respect to CVD (Table 1), Binti et al. compared the mean protein carbonyl levels of patients with acute coronary syndrome, comprising unstable angina, non-ST elevation, and ST-elevation myocardial infarction, with those of the control group. The difference between the groups was notable: 1.63 ± 1.06 nmol/mg in the control group and 3.16 ± 1.29 nmol/mg in the ACS group ($p < 0.0001$) [74]. Gryszczyńska et al. [75] assessed the levels of carbonylated proteins (CP) and advanced oxidation protein products (AOPPs) in patients with abdominal aortic aneurysms (AAAs), aortoiliac occlusive disease (AIOD), and chronic kidney disease (CKD) (predialysis and hemodialysis). The results revealed that the AOPP concentration was highest in the prevalent AAA group, followed by the AIOD group, but lowest in the predialysis and hemodialysis groups. However, CP was greater in the predialysis group than in the AAA or AIOD groups. It is known that increased oxidative stress can lead to CKD, which shares a common pathogenesis mechanism with CVD. This interrelationship has been observed in geriatric patients (aged 60.9 ± 15.2 years) with CKD stages 1–5, where the level of plasma protein carbonyls increases as renal function decreases (as measured by creatinine clearance) ($r = -0.692$, $p < 0.0001$) [76].

5.2 Oxidized Low-Density Lipoprotein

The potential of oxidized low-density lipoproteins (oxLDLs) as biomarkers in CVD has been reviewed in many studies [77–79]. As biomarkers, the most investigated oxLDL component is oxidized phospholipids (oxPLs), and their elevation is thought to play a significant role in oxLDL-induced vascular inflammation and subsequent coronary, carotid and femoral artery diseases [77, 80–82]. OxLDLs, as well as oxPLs, are directly identified by some TLRs (TLR4, TLR6) [83], complement components, and scavenger receptors (CD36) [84–87]. CD36 induces OxLDL uptake and promotes the intracellular formation of cholesterol crystals, further activating the inflammasome via the proinflammatory activity of IL-1 [88].

Furthermore, exogenous oxPLs cause apoptotic cell death, a critical mechanism of atherogenesis [89]. Dijk et al. evaluated six atherosclerotic lesion types of post-mortem carotid endarterectomy by immunostaining for the detection of oxPLs, malondialdehyde (MDA) and apoprotein an (apo(a)) epitopes [85]. The results demonstrated that all atherosclerotic lesions, such as necrotic cores, fibrous caps, foamy macrophages, and VSMCs, expressed oxidation-specific epitopes [85]. Furthermore, Tsimikas et al. established a strong positive correlation between oxPLs/apoB and lipoprotein a (LP(a)) ($r = 0.85$, $p < 0.001$), a cardiovascular risk factor [90]. A meta-analysis of three studies [91], in which two assessed oxLDL levels in participants with HIV (human immunodeficiency virus) disease either with or without associated CVD (468 vs. 487, respectively), revealed that increased oxLDL levels were significantly related to CVD.

Notably, the interaction of oxPLs and plasminogen is related to the increased potential to induce fibrinolysis and thus is associated with a lower atherothrombotic risk [92]. Notably, plasma oxLDL has been shown to be steadily increased in patients with CVD, insulin resistance, diabetes and obesity, regardless of the assay used [16, 78].

5.3 Trans-4-hydroxy-2-nominal and malondialdehyde

Aldehydes such as *trans*-4-hydroxy-2-nominal (4-HNE) and malondialdehyde (MDA) are lipid peroxidation products characterized by their rapid reactivity with proteins to form Michael adducts (advanced lipoxidation end products) [16, 93, 94]. 4-HNE is the most plentiful lipid-derived reactive carbonyl species. It is a major toxic product that induces apoptotic cell death [95]. However, the antioxidant system functions normally, and the cell can degrade modified proteins. In that case, glutathione transferases rapidly neutralize these compounds, especially glutathione-S-transferases 4–4, heme oxygenases, aldehyde dehydrogenases and glutamate-cysteine ligases [96–98]. As mentioned above, proteins participating in reactions with aldehydes are residues of apolipoprotein B (apoB) [99], and subsequent alterations in its structure by MDA increase its affinity for the scavenger receptors of macrophages and cause their transformation into foam cells [100]. 4-HNE provokes cellular oxidative stress in addition to inducing the activation of endoplasmic reticulum stress [101]. Both aldehydes and oxidized phospholipids can trigger inflammation by stimulating the expression of inflammation-related genes [85]. This can lead to an increase in the production of *class A* scavenger receptors on macrophages and smooth muscle cells, as well as strong upregulation of the cytokine TGF- β 1 and activated NF- κ B [88–90]. Notably, 4-HNE, especially MDA and other lipid oxidation end products, are among the most researched and are most commonly used as

oxidative stress markers [77, 93, 102]. Considering CVD, HNE and MDA as markers are preferable to other lipid oxidation products for estimating the risk of acute cardiovascular events, especially myocardial infarction and ischemic stroke [77, 103]. A large study [104] compared the levels of protein-bound HNE products (HNEp) in 61 heart failure (HF) patients with those in 71 healthy individuals. In addition to HNEp, the levels of different types of circulating fatty acids, including n-6 PUFAs, such as linoleic acid, which can conceivably lead to the formation of HNEp, were estimated. In HF, increased HNEp contributed to more severe HF and decreased HDL-C levels. Another work by Taty Zau et al. [105] assessed the effectiveness of a cardiac rehabilitation program in managing systemic oxidative stress in patients with chronic stable coronary disease who underwent coronary artery bypass grafting. A significant and progressive decrease in the oxidative markers of lipid damage, which included MDA and protein carbonyl levels, was observed in this cohort. Additionally, there was an ensuing decrease in superoxide dismutase, catalase, and glutathione peroxidase activities.

5.4 F₂-Isoprostanes

F₂-isoprostanes, which arise from polyunsaturated fatty acid (PUFA) peroxidation, are prostaglandin-like compounds characterized by platelet-activating and vasoconstrictive properties [102, 106, 107]. Both F₂-isoprostanes (F₂-IsoPs) and reactive γ -ketoaldehydes (isolevuglandins) are formed during the nonenzymatic rearrangement of H₂-isoprostanes [16]. The latter compounds are products of the oxidation of arachidonic acid, which is physiologically esterified in tissue phospholipids [108]. Specifically, F₂-IsoPs are frequently treated as the most credible markers for monitoring oxidative stress in vivo because they are correlated with the extent of CVD, reliable outcome prediction and chemical stability [102, 109, 110]. Several studies have established the level of F₂-IsoPs as one of the many risk factors for coronary heart disease (CHD) [111, 112]. Shishehbor et al. quantified nine distinct lipid peroxidation products in the plasma of patients via mass spectrometry. After this, patients were evaluated by diagnostic.

Coronary angiography revealed a statistically significant correlation between higher lipoxidation product levels and CHD. The results of the study revealed that F₂-IsoPs were significantly greater in those diagnosed with CHD (OR 9.7 in the highest F₂-IsoP quartile and plasma levels 1, fivefold greater in CHD) [112]. A subsequent study revealed 93 patients with CHD and 93 healthy controls were confirmed by measuring the levels of F₂-IsoPs along with standard risk markers such as hypercholesterolemia, low HDL, diabetes, body mass

index, systolic blood pressure, CRP, and smoking status [111]. A correlation between higher F₂-IsoP values and a broader spectrum of risk factors was verified, such as between higher F₂-IsoP levels and CHD (OR 27.3 in the highest F₂-IsoP tertile) [111]. Many studies have shown that the levels of F₂-IsoPs can be used to indicate CHD severity. Vassalle et al. reported a relationship between elevated plasma levels of F₂-IsoPs and a greater number of diseased vessels (F₂-IsoP plasma levels are 1.5-fold greater with 1-vessel disease and 2.0-fold greater with multiple vessel disease) [115]. In an extensive systematic review [116], higher levels of plasma F₂-isoprostanes were measured in ischemic stroke patients than in healthy participants. Furthermore, more elevated urinary 8-iso-PGF₂ α (a major F₂-isoprostane isomer) was observed in patients with chronic lower limb ischemia than in healthy controls. However, F₂-isoprostanes were not associated with coronary artery disease.

6 Cardioprotective Drugs with Antioxidant Effects

Current antidiabetic cardiovascular drugs, such as sodium-glucose transport protein 2 inhibitors (SGLT2i), glucagon-like peptide-1 (GLP-1) analogs, and mineralocorticoid receptor antagonists, effectively reduce CVD risk by inhibiting inflammatory and oxidative stress mechanisms [3].

On the basis of numerous clinical trials, the European Society of Cardiology confirmed updated acute and chronic heart failure diagnosis and treatment guidelines, which recommend SGLT2 inhibitors as agents that reduce cardiovascular death, worsening heart failure and hospitalization due to heart failure [117]. The impact of SGLT2i on ameliorating thromboinflammation has been investigated in many basic and clinical studies. Agents such as empagliflozin and ipragliflozin attenuate ROS, VCAM-1 and ICAM-1 in the abdominal aorta of mice [118]. Furthermore, empagliflozin was revealed to decrease mitochondrial production of ROS in the endothelial cells of diabetic and hypertensive elderly patients [119]. Uthman et al. conducted a study to investigate whether empagliflozin and dapagliflozin decrease TNF- α -induced inflammation in human coronary arterial endothelial cells. The results revealed that SGLT2i inhibited ROS generation and thus diminished inflammation in TNF- α -induced coronary arterial endothelial cells [120]. Another study demonstrated that uremic serum from patients with chronic kidney disease harms cardiac microvascular endothelial control of cardiomyocytes and that empagliflozin recovers this intercellular crosstalk by reducing ROS and restoring NO levels in cardiomyocytes, improving their relaxation and contraction [121].

Another antidiabetic agent, GLP-1, is currently being extensively researched. Different clinical trials and

studies have revealed reduced major cardiovascular events in patients treated with GLP-1. In the clinical trial LEADER (Effects of Liraglutide on Cardiovascular Outcomes in Patients With Diabetes With or Without Heart Failure), patients treated with liraglutide had a lower risk of cardiovascular death, AMI, or stroke [122]. GLP-1, as well as SGLT2i, has antioxidant effects by decreasing ROS in endothelial cells, reducing the accumulation of macrophages in the vascular wall and the expression of VCAM-1, ICAM-1, and E-selectins and thus preventing atherosclerotic plaque formation [123]. The activation of the mineralocorticoid receptor (MR) in various cell types plays a crucial role in the development of cardiac hypertrophy and dysfunction, ultimately leading to heart failure [124]. For example, Rac1, a Rho family of GTPase members, acts as a cellular modulator that can activate the MR. In mice undergoing transverse aortic constriction, the activation of Rac1 leads to increased accumulation of MR in the nucleus and increased expression of MR target genes, such as the NOX4 gene, resulting in overproduction of ROS [125]. In rodent models of heart failure, MR antagonists have been shown to reduce cardiac hypertrophy and dysfunction [125]. For example, eplerenone has been shown to decrease myocardial fibrosis and apoptosis, whereas spironolactone inhibits cardiac fibroblast proliferation. Compared with eplerenone, finerenone has been demonstrated to significantly reduce left ventricular wall thickness and mass [126]. The Randomized Aldactone Evaluation Study in 1999 reported a significant reduction in mortality in the spironolactone group [127]. For more than 10 years, eplerenone treatment was associated with a reduction in deaths from CVD or hospitalization for heart failure [128]. MR expression is upregulated in the postinfarct state [129]. In rats with myocardial infarction, there is impaired diastolic function and increased collagen content in the LV interstitium and the aorta [130]. MR antagonists were found to reduce the infarct area and abnormal LV remodeling. In addition to improved left ventricular compliance and elastance, treatment with finerenone reduces interstitial fibrosis in mice with MI [131]. A clinical trial called the Role of eplerenone in Acute Myocardial Infarction–Double-Blind, Early Treatment Initiation, Randomized, placebo-controlled, multicenter study (REMINDER) documented a significant reduction in brain natriuretic peptide (BNP)/N-terminal pro-b-type natriuretic peptide (NT-proBNP) levels and the composite primary endpoint in patients receiving eplerenone within 24 h after ST-elevated myocardial infarction (STEMI) [132].

7 Conclusions

Redox imbalance contributes to oxidative stress and triggers the development and acceleration of CVD. Importantly, excess ROS production leads to endothelial dysfunction, which affects cardiovascular homeostasis and orchestrates thromboinflammation. Oxidative stress biomarkers could be utilized for estimating CVD risk or improving the diagnosis of CVD. To date, only some of the researched biomarkers have been used regularly in the clinic as oxidized low-density lipoproteins because of their unstable nature, hardly detectable levels, or methodological challenges. Nevertheless, targeting ROS generation or using cardioprotective drugs with antioxidant effects might aid in restoring endothelial cell function and improving CVD symptoms. Hence, further discoveries on oxidative stress, its biomarkers, and antioxidants will change the routine clinical approach to CVD treatment.

Abbreviations

CVD _s	Cardiovascular diseases
CHD	Chronic heart disease
EC _s	Endothelial cells
ROS	Reactive oxygen species
NOX _s	Nicotinamide adenine dinucleotide phosphate oxidases
SOD	Superoxide dismutase
NET	Neutrophil extracellular trap
NO	Nitric oxide
vWF	von Willebrand factor
DNA	Deoxyribonucleotide acid
AMI	Acute myocardial infarction
HDL	High-density lipoprotein
VTE	Venous thromboembolism
PUFA	Polyunsaturated fatty acid
4-HNE	Trans-4-hydroxy-2-nominal
MDA	: Malondialdehyde
ApoB	Apolipoprotein B
MMPs	Matrix metalloproteinases
ANG II	Angiotensin II
CRP	C-reactive protein
VSMCs	Vascular smooth muscle cells
oxLDL	Oxidized low-density lipoprotein
LOX-1	Lectin oxidized LDL receptor-1
TNF-α	Tumor necrosis factor alpha
PGI ₂	Prostacyclin
eNOS	Nitric oxide synthase
BH4	Tetrahydrobiopterin
MAPK _s	Mitogen-activated protein kinases
IL-1	Interleukin 1
IL-6	Interleukin-6
VCAM	1-Vascular cell-adhesion molecule-1
ICAM	1-Intracellular cell-adhesion molecule-1
NF-κB	Nuclear factor κB
IL-1β	Interleukin 1β
MHC	Major histocompatibility complex
IFN-γ	Interferon-γ

Acknowledgements

Not applicable.

Author Contributions

Writing—original draft preparation: Julija Valaitienė. Writing—review and editing: Agnė Laučytė-Cibulskienė.

Funding

Open access funding provided by Lund University. Faculty of Medicine, Lund University, Sweden.

Availability of Data and Materials

All data pertaining to this manuscript are freely available.

Declarations**Conflict of Interest**

All the authors certify that they have no affiliations with or involvement in any organization or entity with any financial or nonfinancial interest in the subject matter or materials discussed in this manuscript.

Ethical Approval and Consent to Participate

Not applicable.

Consent for Publication

Not applicable.

Received: 7 March 2024 Accepted: 15 October 2024

Published online: 29 October 2024

References

- Roth GA, Abate D, Abate KH, Abay SM, Abbafati C, Abbasi N, et al. Global, regional, and national age-sex-specific mortality for 282 causes of death in 195 countries and territories, 1980–2017: a systematic analysis for the Global Burden of Disease Study 2017. *The Lancet*. 2018;392(10159):1736–88.
- Benjamin EJ, Muntner P, et al. Heart disease and stroke statistics—2019 Update: A Report From the American Heart Association. *Circulation*. 2019. <https://doi.org/10.1161/CIR.0000000000000659>.
- Steven S, Frenis K, Oelze M, Kalinovic S, Kuntic M, Bayo Jimenez MT, et al. Vascular inflammation and oxidative stress: major triggers for cardiovascular disease. *Oxid Med Cell Longev*. 2019;2019: e7092151.
- Burtenshaw D, Hakimjavadi R, Redmond EM, Cahill PA. Nox, reactive oxygen species and regulation of vascular cell fate. *Antioxidants*. 2017;6(4):90.
- Xu S, Ilyas I, Little PJ, Li H, Kamato D, Zheng X, et al. Endothelial dysfunction in atherosclerotic cardiovascular diseases and beyond: from mechanism to pharmacotherapies. *Pharmacol Rev*. 2021;73(3):924–67.
- Al-Shehri SS. Reactive oxygen and nitrogen species and innate immune response. *Biochimie*. 2021;181:52–64.
- Ito H, Kurokawa H, Matsui H. Mitochondrial reactive oxygen species and heme, non-heme iron metabolism. *Arch Biochem Biophys*. 2021;700: 108695.
- Brandes RP, Weissmann N, Schröder K. Nox family NADPH oxidases: molecular mechanisms of activation. *Free Radic Biol Med*. 2014;76:208–26.
- Granger DN, Kvietys PR. Reperfusion injury and reactive oxygen species: the evolution of a concept. *Redox Biol*. 2015;6:524–51.
- Zhang B, Pan C, Feng C, Yan C, Yu Y, Chen Z, et al. Role of mitochondrial reactive oxygen species in homeostasis regulation. *Redox Rep Commun Free Radic Res*. 2022;27(1):45–52.
- Kurokawa H, Ito H, Matsui H. Monascus purpureus induced apoptosis on gastric cancer cell by scavenging mitochondrial reactive oxygen species. *J Clin Biochem Nutr*. 2017;61(3):189–95.
- Wallert M, Ziegler M, Wang X, Maluenda A, Xu X, Yap ML, et al. α -Tocopherol preserves cardiac function by reducing oxidative stress and inflammation in ischemia/reperfusion injury. *Redox Biol*. 2019;26: 101292.
- Fu A, van Rooyen L, Evans L, Armstrong N, Avizonis D, Kin T, et al. Glucose metabolism and pyruvate carboxylase enhance glutathione synthesis and restrict oxidative stress in pancreatic islets. *Cell Rep*. 2021;37(8): 110037.
- Kim JL, Reader BF, Dumond C, Lee Y, Mokadam NA, Black SM, et al. Pegylated-catalase is protective in lung ischemic injury and oxidative stress. *Ann Thorac Surg*. 2021;111(3):1019–27.
- Wert KJ, Velez G, Cross MR, Wagner BA, Teoh-Fitzgerald ML, Buettner GR, et al. Extracellular superoxide dismutase (SOD3) regulates oxidative stress at the vitreoretinal interface. *Free Radic Biol Med*. 2018;124:408–19.
- Frijhoff J, Winyard PG, Zarkovic N, Davies SS, Stocker R, Cheng D, et al. Clinical relevance of biomarkers of oxidative stress. *Antioxid Redox Signal*. 2015;23(14):1144–70.
- Califf RM. Biomarker definitions and their applications. *Exp Biol Med*. 2018;243(3):213–21.
- Ghosh A, Gao L, Thakur A, Siu PM, Lai CWK. Role of free fatty acids in endothelial dysfunction. *J Biomed Sci*. 2017;24(1):50.
- Shaito A, Aramouni K, Assaf R, Parenti A, Orekhov A, Yazbi AE, et al. Oxidative stress-induced endothelial dysfunction in cardiovascular diseases. *Front Biosci-Landmark*. 2022;27(3):0105.
- Capurso C, Capurso A. From excess adiposity to insulin resistance: the role of free fatty acids. *Vascul Pharmacol*. 2012;57(2):91–7.
- Giacco F, Brownlee M, Schmidt AM. Oxidative stress and diabetic complications. *Circ Res*. 2010;107(9):1058–70.
- Rajendran P, Rengarajan T, Thangavel J, Nishigaki Y, Sakthisekaran D, Sethi G, et al. The vascular endothelium and human diseases. *Int J Biol Sci*. 2013;9(10):1057–69.
- Vanhoutte PM, Shimokawa H, Tang EHC, Feletou M. Endothelial dysfunction and vascular disease. *Acta Physiol*. 2009;196(2):193–222.
- Förstermann U. Nitric oxide and oxidative stress in vascular disease. *Pflug Arch - Eur J Physiol*. 2010;459(6):923–39.
- Zorov DB, Juhaszova M, Sollott SJ. Mitochondrial reactive oxygen species (ROS) and ROS-induced ROS release. *Physiol Rev*. 2014;94(3):909–50.
- Kaarniranta K, Pawlowska E, Szczepanska J, Jablowska A, Blasiak J. Role of mitochondrial DNA damage in ROS-mediated pathogenesis of age-related macular degeneration (AMD). *Int J Mol Sci*. 2019;20(10):2374.
- Davidson SM, Duchon MR. Endothelial mitochondria. *Circ Res*. 2007;100(8):1128–41.
- Aggarwal V, Tuli HS, Varol A, Thakral F, Yerer MB, Sak K, et al. Role of reactive oxygen species in cancer progression: molecular mechanisms and recent advancements. *Biomolecules*. 2019;9(11):735.
- Mitra S, Goyal T, Mehta JL. Oxidized LDL, LOX-1 and atherosclerosis. *Cardiovasc Drugs Ther*. 2011;25(5):419–29.
- Giró O, Jiménez A, Pané A, Badimon L, Ortega E, Chiva-Blanch G. Extracellular vesicles in atherothrombosis and cardiovascular disease: friends and foes. *Atherosclerosis*. 2021;330:61–75.
- Zhang J. Biomarkers of endothelial activation and dysfunction in cardiovascular diseases. *Rev Cardiovasc Med*. 2022;23(2):73.
- Bennett MR, Sinha S, Owens GK. Vascular smooth muscle cells in atherosclerosis. *Circ Res*. 2016;118(4):692–702.
- Yoshida T, Gan Q, Owens GK. Krüppel-like factor 4, Elk-1, and histone deacetylases cooperatively suppress smooth muscle cell differentiation markers in response to oxidized phospholipids. *Am J Physiol-Cell Physiol*. 2008;295(5):C1175–82.
- Mandel J, Casari M, Stepanyan M, Martyanov A, Deppermann C. Beyond hemostasis: platelet innate immune interactions and thromboinflammation. *Int J Mol Sci*. 2022;23(7):3868.
- Kasper B, Brandt E, Bulfone-Paus S, Petersen F. Platelet factor 4 (PF-4)-induced neutrophil adhesion is controlled by src-kinases, whereas PF-4-mediated exocytosis requires the additional activation of p38 MAP kinase and phosphatidylinositol 3-kinase. *Blood*. 2004;103(5):1602–10.
- Mostafa MN, Osama M. The implications of neutrophil extracellular traps in the pathophysiology of atherosclerosis and atherothrombosis. *Exp Biol Med Maywood NJ*. 2020;245(15):1376–84.
- Ionita MG, van den Borne P, Catanzariti LM, Moll FL, de Vries JPPM, Pasterkamp G, et al. High neutrophil numbers in human carotid atherosclerotic plaques are associated with characteristics of rupture-prone lesions. *Arterioscler Thromb Vasc Biol*. 2010;30(9):1842–8.
- Moore K, Sheedy F, Fisher E. Macrophages in atherosclerosis: a dynamic balance. *Nat Rev Immunol*. 2013;13(10):709–21.

39. Roy P, Orecchioni M, Ley K. How the immune system shapes atherosclerosis: roles of innate and adaptive immunity. *Nat Rev Immunol*. 2022;22(4):251–65.
40. Zernecke A. Dendritic cells in atherosclerosis. *Arterioscler Thromb Vasc Biol*. 2015;35(4):763–70.
41. Galkina E, Kadl A, Sanders J, Varughese D, Sarembock IJ, Ley K. Lymphocyte recruitment into the aortic wall before and during development of atherosclerosis is partially L-selectin dependent. *J Exp Med*. 2006;203(5):1273–82.
42. Kloc M, Kubiak JZ, Ghobrial RM. Macrophage-, dendritic-, smooth muscle-, endothelium-, and stem cells-derived foam cells in atherosclerosis. *Int J Mol Sci*. 2022;23(22):14154.
43. Vallejo J, Cochain C, Zernecke A, Ley K. Heterogeneity of immune cells in human atherosclerosis revealed by scRNA-Seq. *Cardiovasc Res*. 2021;117(13):2537–43.
44. Senoner T, Dichtl W. Oxidative stress in cardiovascular diseases: still a therapeutic target? *Nutrients*. 2019;11(9):2090.
45. Kattoor AJ, Pothineni NVK, Palagiri D, Mehta JL. Oxidative stress in atherosclerosis. *Curr Atheroscler Rep*. 2017;19(11):42.
46. Meo SD, Venditti P. Evolution of the knowledge of free radicals and other oxidants. *Oxid Med Cell Longev*. 2020. <https://doi.org/10.1155/2020/9829176>.
47. He L, He T, Farrar S, Ji L, Liu T, Ma X. Antioxidants maintain cellular redox homeostasis by elimination of reactive oxygen species. *Cell Physiol Biochem*. 2017;44(2):532–53.
48. Burtenshaw D, Hakimjavadi R, Redmond EM, Cahill PA. Nox, reactive oxygen species and regulation of vascular cell fate. *Antioxidants*. 2017;6(4):90.
49. Sinha K, Das J, Pal PB, Sil PC. Oxidative stress: the mitochondria-dependent and mitochondria-independent pathways of apoptosis. *Arch Toxicol*. 2013;87(7):1157–80.
50. Donato AJ, Morgan RG, Walker AE, Lesniewski LA. Cellular and molecular biology of aging endothelial cells. *J Mol Cell Cardiol*. 2015;89:122–35.
51. Chen X, Zhang T, Du G. Advanced glycation end products serve as ligands for lectin-like oxidized low-density lipoprotein receptor-1 (LOX-1): biochemical and binding characterizations assay. *Cell Biochem Funct*. 2008;26(7):760–70.
52. Jono T, Miyazaki A, Nagai R, Sawamura T, Kitamura T, Horiuchi S. Lectin-like oxidized low density lipoprotein receptor-1 (LOX-1) serves as an endothelial receptor for advanced glycation end products (AGE). *FEBS Lett*. 2002;511(1–3):170–4.
53. Taye A, Sawamura T, Morawietz H. Aldosterone augments LOX-1-mediated low-density lipoprotein uptake in human umbilical artery endothelial cells. *Pharmacol Rep*. 2010;62(2):311–8.
54. Sankaralingam S, Xu Y, Sawamura T, Davidge ST. Increased lectin-like oxidized low-density lipoprotein receptor-1 expression in the maternal vasculature of women with preeclampsia. *Hypertension*. 2009;53(2):270–7.
55. Touyz RM, Briones AM. Reactive oxygen species and vascular biology: implications in human hypertension. *Hypertens Res*. 2011;34(1):5–14.
56. Shah PK. Mechanisms of plaque vulnerability and rupture. *J Am Coll Cardiol*. 2003;41(4, Supplement):S15–22.
57. Asada Y, Yamashita A, Sato Y, Hatakeyama K. Pathophysiology of atherothrombosis: mechanisms of thrombus formation on disrupted atherosclerotic plaques. *Pathol Int*. 2020;70(6):309–22.
58. Libby P. Mechanisms of acute coronary syndromes and their implications for therapy. *N Engl J Med*. 2013;368(21):2004–13.
59. Cimini M, Kishore R. Role of podoplanin-positive cells in cardiac fibrosis and angiogenesis after ischemia. *Front Physiol*. 2021. <https://doi.org/10.3389/fphys.2021.667278>.
60. Inoue O, Hokamura K, Shirai T, Osada M, Tsukiji N, Hatakeyama K, et al. Vascular smooth muscle cells stimulate platelets and facilitate thrombus formation through platelet CLEC-2: implications in atherothrombosis. *PLoS ONE*. 2015;10(9):e0139357.
61. Sato Y, Hatakeyama K, Yamashita A, Marutsuka K, Sumiyoshi A, Asada Y. Proportion of fibrin and platelets differs in thrombi on ruptured and eroded coronary atherosclerotic plaques in humans. *Heart*. 2005;91(4):526–30.
62. Bogdanov VY, Versteeg HH. “Soluble tissue factor” in the 21st century: definitions, biochemistry, and pathophysiological role in thrombus formation. *Semin Thromb Hemost*. 2015;41:700–7.
63. Jamasbi J, Ayabe K, Goto S, Nieswandt B, Peter K, Siess W. Platelet receptors as therapeutic targets: past, present and future. *Thromb Haemost*. 2017;117(07):1249–57.
64. Badimon L, Vilahur G, Rocca B, Patrono C. The key contribution of platelet and vascular arachidonic acid metabolism to the pathophysiology of atherothrombosis. *Cardiovasc Res*. 2021;117(9):2001–15.
65. Akagawa M. Protein carbonylation: molecular mechanisms, biological implications, and analytical approaches. *Free Radic Res*. 2021;55(4):307–20.
66. Dainin K, Ide R, Maeda A, Suyama K, Akagawa M. Pyridoxamine scavenges protein carbonyls and inhibits protein aggregation in oxidative stress-induced human HepG2 hepatocytes. *Biochem Biophys Res Commun*. 2017;486(3):845–51.
67. Estévez M. Protein carbonyls in meat systems: a review. *Meat Sci*. 2011;89(3):259–79.
68. Stoner L, Lucero AA, Palmer BR, Jones LM, Young JM, Faulkner J. Inflammatory biomarkers for predicting cardiovascular disease. *Clin Biochem*. 2013;46(15):1353–71.
69. Yan SF, Ramasamy R, Naka Y, Schmidt AM. Glycation, inflammation, and RAGE: a scaffold for the macrovascular complications of diabetes and beyond. *Circ Res*. 2003;93(12):1159–69.
70. Greilberger J, Koidl C, Greilberger M, Lamprecht M, Schroecksnadel K, Leblhuber F, et al. Malondialdehyde, carbonyl proteins and albumin-disulphide as useful oxidative markers in mild cognitive impairment and Alzheimer’s disease. *Free Radic Res*. 2008;42(7):633–8.
71. Henning C, Glomb MA. Pathways of the Maillard reaction under physiological conditions. *Glycoconj J*. 2016;33(4):499–512.
72. Miyata T, Ueda Y, Shinzato T, Iida Y, Tanaka S, Kurokawa K, et al. Accumulation of albumin-linked and free-form pentosidine in the circulation of uremic patients with end-stage renal failure: renal implications in the pathophysiology of pentosidine. *J Am Soc Nephrol*. 1996;7(8):1198.
73. Dalle-Donne I, Rossi R, Giustarini D, Milzani A, Colombo R. Protein carbonyl groups as biomarkers of oxidative stress. *Clin Chim Acta*. 2003;329(1):23–38.
74. Binti NN, Ferdausi N, Anik MdEK, Islam LN. Association of albumin, fibrinogen, and modified proteins with acute coronary syndrome. *PLoS ONE*. 2022;17(7):e0271882.
75. Gryszczyńska B, Formanowicz D, Budzyń M, Wanik-Kossowska M, Pawliczak E, Formanowicz P, et al. Advanced oxidation protein products and carbonylated proteins as biomarkers of oxidative stress in selected atherosclerosis-mediated diseases. *BioMed Res Int*. 2017;2017:4975264.
76. Matsuyama Y, Terawaki H, Terada T, Era S. Albumin thiol oxidation and serum protein carbonyl formation are progressively enhanced with advancing stages of chronic kidney disease. *Clin Exp Nephrol*. 2009;13(4):308–15.
77. Negre-Salvayre A, Auge N, Ayala V, Basaga H, Boada J, Brenke R, et al. Pathological aspects of lipid peroxidation. *Free Radic Res*. 2010;44(10):1125–71.
78. Trpkovic A, Resanovic I, Stanimirovic J, Radak D, Mousa SA, Cenic-Milosevic D, et al. Oxidized low-density lipoprotein as a biomarker of cardiovascular diseases. *Crit Rev Clin Lab Sci*. 2015;52(2):70–85.
79. Tsimikas S. Measures of oxidative stress. *Clin Lab Med*. 2006;26(3):571–90.
80. Freigang S. The regulation of inflammation by oxidized phospholipids. *Eur J Immunol*. 2016;46(8):1818–25.
81. Frostegård J. Low level natural antibodies against phosphorylcholine: a novel risk marker and potential mechanism in atherosclerosis and cardiovascular disease. *Clin Immunol*. 2010;134(1):47–54.
82. Ravandi A, Leibundgut G, Hung MY, Patel M, Hutchins PM, Murphy RC, et al. Release and capture of bioactive oxidized phospholipids and oxidized cholesteryl esters during percutaneous coronary and peripheral arterial interventions in humans. *J Am Coll Cardiol*. 2014;63(19):1961–71.
83. Imai Y, Kuba K, Neely GG, Yaghubian-Malhami R, Perkmann T, van Loo G, et al. Identification of oxidative stress and toll-like receptor 4 signaling as a key pathway of acute lung injury. *Cell*. 2008;133(2):235–49.
84. Binder CJ, Papac-Milicevic N, Witztum JL. Innate sensing of oxidation-specific epitopes in health and disease. *Nat Rev Immunol*. 2016;16(8):485–97.
85. van Dijk RA, Kolodgie F, Ravandi A, Leibundgut G, Hu PP, Prasad A, et al. Differential expression of oxidation-specific epitopes and

- apolipoprotein(a) in progressing and ruptured human coronary and carotid atherosclerotic lesions. *J Lipid Res.* 2012;53(12):2773–90.
86. Miller YI, Choi SH, Wiesner P, Fang L, Harkewicz R, Hartvigsen K, et al. Oxidation-specific epitopes are danger-associated molecular patterns recognized by pattern recognition receptors of innate immunity. *Circ Res.* 2011;108(2):235–48.
 87. Podrez EA, Poliakov E, Shen Z, Zhang R, Deng Y, Sun M, et al. Identification of a novel family of oxidized phospholipids that serve as ligands for the macrophage scavenger receptor CD36 *. *J Biol Chem.* 2002;277(41):38503–16.
 88. Kadl A, Sharma PR, Chen W, Agrawal R, Meher AK, Rudraiah S, et al. Oxidized phospholipid-induced inflammation is mediated by Toll-like receptor 2. *Free Radic Biol Med.* 2011;51(10):1903–9.
 89. Chen R, Yang L, McIntyre TM. Cytotoxic phospholipid oxidation products. *J Biol Chem.* 2007;282(34):24842–50.
 90. Tsimikas S, Clopton P, Brilakis ES, Marcovina SM, Khera A, Miller ER, et al. Relationship of oxidized phospholipids on apolipoprotein B-100 particles to race/ethnicity, apolipoprotein(a) isoform size, and cardiovascular risk factors. *Circulation.* 2009;119(13):1711–9.
 91. Hong CG, Florida E, Li H, Parel PM, Mehta NN, Sorokin AV. Oxidized low-density lipoprotein associates with cardiovascular disease by a vicious cycle of atherosclerosis and inflammation: a systematic review and meta-analysis. *Front Cardiovasc Med.* 2023. <https://doi.org/10.3389/fcvm.2022.1023651>.
 92. Leibundgut G, Arai K, Orsoni A, Yin H, Scipione C, Miller ER, et al. Oxidized phospholipids are present on plasminogen, affect fibrinolysis, and increase following acute myocardial infarction. *J Am Coll Cardiol.* 2012;59(16):1426–37.
 93. Gianazza E, Brioschi M, Fernandez AM, Banfi C. Lipoxidation in cardiovascular diseases. *Redox Biol.* 2019;23: 101119.
 94. Spickett CM. The lipid peroxidation product 4-hydroxy-2-nonenal: advances in chemistry and analysis. *Redox Biol.* 2013;1(1):145–52.
 95. Schaur RJ. Basic aspects of the biochemical reactivity of 4-hydroxynonenal. *Mol Aspects Med.* 2003;24(4):149–59.
 96. Balogh LM, Atkins WM. Interactions of glutathione transferases with 4-hydroxynonenal. *Drug Metab Rev.* 2011;43(2):165–78.
 97. Esterbauer H. Cytotoxicity and genotoxicity of lipid-oxidation products. *Am J Clin Nutr.* 1993;57(5):779S–786S.
 98. Zhang H, Forman HJ. Signaling pathways involved in phase II gene induction by α , β -unsaturated aldehydes. *Toxicol Ind Health.* 2009;25(4–5):269–78.
 99. Leibundgut G, Witztum JL, Tsimikas S. Oxidation-specific epitopes and immunological responses: translational biotherapeutic implications for atherosclerosis. *Curr Opin Pharmacol.* 2013;13(2):168–79.
 100. Steinbrecher U. Receptors for oxidized low density lipoprotein. *Biochim Biophys Acta BBA - Mol Cell Biol Lipids.* 1999;1436(3):279–98.
 101. Sanson M, Augé N, Vindis C, Muller C, Bando Y, Thiers JC, et al. Oxidized low-density lipoproteins trigger endoplasmic reticulum stress in vascular cells. *Circ Res.* 2009;104(3):328–36.
 102. Gaggini M, Sabatino L, Vassalle C. Conventional and innovative methods to assess oxidative stress biomarkers in the clinical cardiovascular setting. *Biotechniques.* 2020;68(4):223–31.
 103. Tsimikas S. Oxidized low-density lipoprotein biomarkers in atherosclerosis. *Curr Atheroscler Rep.* 2006;8(1):55–61.
 104. Asselin C, Ducharme A, Ntimbane T, Ruiz M, Fortier A, Guertin MC, et al. Circulating levels of linoleic acid and HDL-cholesterol are major determinants of 4-hydroxynonenal protein adducts in patients with heart failure. *Redox Biol.* 2014;2:148–55.
 105. Taty Zau JF, Costa Zeferino R, Sandrine Mota N, Fernandes Martins G, Manoel Serra S, Bonates da Cunha T, et al. Exercise through a cardiac rehabilitation program attenuates oxidative stress in patients submitted to coronary artery bypass grafting*. *Redox Rep Commun Free Radic Res.* 2017;23(1):94–9.
 106. Bauer J, Ripberger A, Frantz S, Ergün S, Schwedhelm E, Benndorf RA. Pathophysiology of isoprostanes in the cardiovascular system: implications of isoprostane-mediated thromboxane A2 receptor activation. *Br J Pharmacol.* 2014;171(13):3115–31.
 107. Galano JM, Mas E, Barden A, Mori TA, Signorini C, De Felice C, et al. Isoprostanes and neuroprostanes: total synthesis, biological activity and biomarkers of oxidative stress in humans. *Prostaglandins Other Lipid Mediat.* 2013;107:95–102.
 108. Roberts LJ, Fessel JP, Davies SS. The biochemistry of the isoprostane, neuroprostane, and isofuran pathways of lipid peroxidation. *Brain Pathol.* 2006;15(2):143–8.
 109. Czerska M, Zieliński M, Gromadzińska J. Isoprostanes – A novel major group of oxidative stress markers. *Int J Occup Med Environ Health.* 2015;29(2):179–90.
 110. Davies SS, Roberts LJ. F2-isoprostanes as an indicator and risk factor for coronary heart disease. *Free Radic Biol Med.* 2011;50(5):559–66.
 111. Schwedhelm E, Bartling A, Lenzen H, Tsikas D, Maas R, Brümmer J, et al. Urinary 8-iso-prostaglandin F2 α as a risk marker in patients with coronary heart disease. *Circulation.* 2004;109(7):843–8.
 112. Shishehbor MH, Zhang R, Medina H, Brennan ML, Brennan DM, Ellis SG, et al. Systemic elevations of free radical oxidation products of arachidonic acid are associated with angiographic evidence of coronary artery disease. *Free Radic Biol Med.* 2006;41(11):1678–83.
 113. Zhang ZJ. Systematic review on the association between F₂-isoprostanes and cardiovascular disease. *Ann Clin Biochem Int J Lab Med.* 2013;50(2):108–14.
 114. Ito T, Fujita H, Tani T, Ohte N. Malondialdehyde-modified low-density lipoprotein is a predictor of cardiac events in patients with stable angina on lipid-lowering therapy after percutaneous coronary intervention using drug-eluting stent. *Atherosclerosis.* 2015;239(2):311–7.
 115. Vassalle C, Petrozzi L, Botto N, Andreassi MG, Zucchelli GC. Oxidative stress and its association with coronary artery disease and different atherogenic risk factors. *J Intern Med.* 2004;256(4):308–15.
 116. Zhang ZJ. Systematic review on the association between F2-isoprostanes and cardiovascular disease. *Ann Clin Biochem.* 2013;50(2):108–14.
 117. (2023) Focused Update of the 2021 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure
 118. Yanai H, Adachi H, Hakoshima M, Katsuyama H. Significance of endothelial dysfunction amelioration for sodium-glucose cotransporter 2 inhibitor-induced improvements in heart failure and chronic kidney disease in diabetic patients. *Metabolites.* 2023;13(6):736.
 119. Mone P, Varzideh F, Jankauskas SS, Pansini A, Lombardi A, Frullone S, et al. SGLT2 inhibition via empagliflozin improves endothelial function and reduces mitochondrial oxidative stress: insights from frail hypertensive and diabetic patients. *Hypertens Dallas Tex.* 2022;79(8):1633–43.
 120. Uthman L, Homayr A, Juni RP, Spin EL, Kerindongo R, Boomsma M, et al. Empagliflozin and dapagliflozin reduce ROS generation and restore NO bioavailability in tumor necrosis factor α -stimulated human coronary arterial endothelial cells. *Cell Physiol Biochem Int J Exp Cell Physiol Biochem Pharmacol.* 2019;53(5):865–86.
 121. Juni RP, Al-Shama R, Kuster DWD, van der Velden J, Hamer HM, Vervloet MG, et al. Empagliflozin restores chronic kidney disease-induced impairment of endothelial regulation of cardiomyocyte relaxation and contraction. *Kidney Int.* 2021;99(5):1088–101.
 122. Marso SP, Baeres FMM, Bain SC, Goldman B, Husain M, Nauck MA, et al. Effects of liraglutide on cardiovascular outcomes in patients with diabetes with or without heart failure. *J Am Coll Cardiol.* 2020;75(10):1128–41.
 123. Gallo G, Volpe M. Potential mechanisms of the protective effects of the cardiometabolic drugs type-2 sodium-glucose transporter inhibitors and glucagon-like peptide-1 receptor agonists in heart failure. *Int J Mol Sci.* 2024;25(5):2484.
 124. Parksook WW, Williams GH. Aldosterone and cardiovascular diseases. *Cardiovasc Res.* 2023;119(1):28–44.
 125. Adam O, Lavall D, Theobald K, Hohl M, Grube M, Ameling S, et al. Rac1-induced connective tissue growth factor regulates connexin 43 and n-cadherin expression in atrial fibrillation. *J Am Coll Cardiol.* 2010;55(5):469–80.
 126. Kuster GM, Kotlyar E, Rude MK, Siwik DA, Liao R, Colucci WS, et al. Mineralocorticoid receptor inhibition ameliorates the transition to myocardial failure and decreases oxidative stress and inflammation in mice with chronic pressure overload. *Circulation.* 2005;111(4):420–7.
 127. Pitt B, Zannad F, Remme WJ, et al. The effect of spironolactone on morbidity and mortality in patients with severe heart failure. randomized aldactone evaluation study investigators. *N Engl J Med.* 1999;341(10):709–17. <https://doi.org/10.1056/NEJM199909023411001>.
 128. Zannad F, McMurray JJV, Krum H, Van Veldhuisen DJ, Swedberg K, Shi H, et al. Eplerenone in patients with systolic heart failure and mild symptoms. *N Engl J Med.* 2011;364(1):11–21.

129. Milik E, Szczepańska-Sadowska E, Maśliński W, Cudnoch-Jedrzejewska A. Enhanced expression of mineralocorticoid receptors in the heart after the myocardial infarct in rats. *J Physiol Pharmacol*. 2007;58(4):745–55.
130. Mihailidou AS, Loan Le TY, Mardini M, Funder JW. Glucocorticoids activate cardiac mineralocorticoid receptors during experimental myocardial infarction. *Hypertension*. 2009;54(6):1306–12.
131. Gueret A, Harouki N, Favre J, et al. Vascular smooth muscle mineralocorticoid receptor contributes to coronary and left ventricular dysfunction after myocardial infarction. *Hypertension*. 2016;67(4):717–23. <https://doi.org/10.1161/HYPERTENSIONAHA.115.06709>.
132. Montalescot G, Pitt B, Lopez De Sa E, Hamm CW, Flather M, Verheugt F, et al. Early eplerenone treatment in patients with acute ST-elevation myocardial infarction without heart failure: The Randomized Double-Blind Reminder Study. *Eur Heart J*. 2014;35(34):2295–302.