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CRISPR-CAS9 GENOME ENGINEERING IN KLUYVEROMYCES MARXIANUS FOR ENHANCED SECRETION OF RECOMBINANT ANTIBODIES

Justina Žičkutė¹, Danguolė Žiogienė¹, Alma Gedvilaitė¹

¹Department of Eukaryote Gene Engineering, Institute of Biotechnology, Life Sciences Center, Vilnius University, Lithuania zickutejustina@gmail.com

Recombinant antibodies (RAbs) are important in diagnostics, research, biotechnology, and therapeutics due to their high specificity, stability, and ease of modification [1]. Yeasts, which are easily genetically modified and cultivated, are often preferred as a cost-effective system for RAbs production. The species Kluyveromyces marxianus is known for its efficient production and secretion of properly folded and active native and recombinant proteins, including RAbs [2]. To improve RAbs production technologies, yeasts can be genetically modified to create mutant strains with enhanced protein secretion properties.

Protein glycosylation, a crucial post-translational modification, involves attaching a glycan to a protein, ensuring its proper folding, activity, and stability. Dolichol kinase (DK), encoded by the essential SEC59 gene, plays a role in glycosylation processes within the endoplasmic reticulum [3]. The reduced activity of DK, along with changes in glycosylation levels and the activity of other proteins in the secretory pathway, may lead to enhanced recombinant protein secretion [4]. Enhanced RAbs secretion can also be achieved by reducing the activity of intracellular peptidases, as RAbs are highly prone to proteolysis and often undergo degradation [5].

The aim of this study was to apply efficient CRISPR-Cas9 genome editing technology to construct a K. marxianus strain that displays improved secretion of RAbs. In this investigation, a K. marxianus yeast WSS-Apep4 strain was created by introducing mutations encoding G418S and I432S changes in the DK amino acids, and by disrupting the gene encoding vacuolar peptidase (PEP4). The introduction of mutations in the DK-encoding gene led to changes in DK activity, as indicated by reduced glycosylation efficiency of carboxypeptidase Y in the WSS strain. Additionally, the disruption of the PEP4 gene in yeast resulted in a decrease in the proteolytic degradation of RAbs. A secretion assay of the single-chain antibody fragment (scFv) linked to an antibody fragment crystallizable Fc (scFv-Fc) against Gardnerella vaginalis vaginolysin was performed and detected in yeast growth medium by Western blot. The results indicated that the constructed K. marxianus WSS-Apep4 strain secreted recombinant scFv-Fc protein more efficiently compared to the wild-type K. marxianus strain. However, the secretion of RAbs in yeast also depends on the specific properties of the recombinant protein, and further studies are necessary. The newly constructed K. marxianus WSS-Apep4 mutant strain could be beneficial for future research aimed at enhancing RAbs production technologies.

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