



Article

# Dietary Fat Intake and Indices of Blood Profiles in High-Performance Athletes: An Exploratory Study Focusing on Platelet Variables

Marius Baranauskas <sup>1,\*</sup>, Ingrida Kupčiūnaitė <sup>1</sup>, Jurgita Lieponienė <sup>1</sup> and Rimantas Stukas <sup>2</sup>

- Faculty of Biomedical Sciences, State Higher Education Institution Panevėžys College, 35200 Panevėžys, Lithuania; ingrida.kupciunaite@panko.lt (I.K.); jurgita.lieponiene@panko.lt (J.L.)
- Department of Public Health, Institute of Health Sciences, Faculty of Medicine, Vilnius University, 01513 Vilnius, Lithuania; rimantas.stukas@mf.vu.lt
- \* Correspondence: baranauskas.marius@panko.lt

#### **Abstract**

**Background/Objectives**: There is a sudden and noticeably increasing focus on naturally found antiplatelet inhibitors that humans can use habitually. Given that athletes receive annual training with periods of recovery that are not always suitably adapted to the workload, this study aimed to explore the association between dietary fat intakes and the indices of blood profiles, concentrating on platelet variables in a sample of highperformance athletes. **Methods**: The sample encompassed 19.8  $\pm$  2.2-year-old Lithuanian high-performance athletes (n = 82). The assessment of the nutritional profile of study participants was performed using a 3-day food record approach. In laboratory settings, the hematology profile of athletes was assessed via the Nihon Khoden automated hematology analyzer. Results: The recorded mean consumption of energy, carbohydrates, protein, and fat in elite athletes was 49 kcal/kg/day, 5.4 g/kg/day, 1.6 g/kg/day, and 40.3% of energy intake (EI), respectively. The study highlighted the excessive consumption of saturated fatty acids (FA) (13.4-14.3% of EI) and dietary cholesterol (698-982 mg/day). Also, considering that the ideal human omega-6 to omega-3 FA ratio is commonly deemed to be between 1:1 and 4:1, an athlete's 'Western diet' was heavily skewed with a ratio fluctuating from 18.9:1 to 19:4 in favor of omega-6 FA. Furthermore, the study found that the outcomes related to slightly higher levels of blood platelet counts and plateletcrit, however, being within normal limits, were associated with a higher intake of omega-6 FA (adjusted odds ratio (AOR) 9.5, 95% confidence interval (CI) 1.2; 9.9, p = 0.029). A higher platelet-to-hemoglobin ratio as a novel indirect blood-based biomarker pronouncing the potential inflammatory processes in the body revealed the reverse relationship of higher intake levels of dietary omega-3 FA (AOR 6.7, 95% CI 1.3; 12.2, p = 0.029), omega-6 FA (AOR 6.2, 95% CI 2.7; 11.5, p = 0.009), and saturated FA (AOR 8.5, 95% CI 1.5; 9.1, p = 0.020) among elite athletes. **Conclusions**: The prospect of personalized nutrition targeted at the professional athletes' segment may provide an innovative opportunity to increase athletes' capacity to manage the platelet function via diet while stressing the importance of further empirical experimental research in this dynamic and vital biomedical field.

**Keywords:** blood profiles; nutritional status; omega-3 fatty acids; platelet variables; public health



Academic Editor: Darryn S. Willoughby

Received: 6 October 2025 Revised: 25 October 2025 Accepted: 28 October 2025 Published: 30 October 2025

Citation: Baranauskas, M.; Kupčiūnaitė, I.; Lieponienė, J.; Stukas, R. Dietary Fat Intake and Indices of Blood Profiles in High-Performance Athletes: An Exploratory Study Focusing on Platelet Variables. Nutrients 2025, 17, 3418. https:// doi.org/10.3390/nu17213418

Copyright: © 2025 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

Nutrients 2025, 17, 3418 2 of 17

# 1. Introduction

It has been well established that dietary habits play a potential role in influencing the indices of blood profiles, including the platelet function, a fundamental component in the blood coagulation cascade [1]. It should be emphasized that thrombocytes, or platelets, are the smallest cell elements in the human blood [2] that play a significant role in supporting the equilibrium of hemostasis [3–6]. Derived from the bone marrow cells that mature into platelets [7], these non-nucleated cell fragments circulate in the blood circulation as precursors capable of responding to vascular damage [8,9]. Upon detecting an injury, platelets join the lesion area, aggregate with each other, and start a cascade of reactions. This results in a hemostatic stopper that effectively prevents further bleeding [10,11]. In addition, platelets are engaged in many other physiological processes, including angiogenesis, immunity, and inflammation, as significant agents under numerous physiological and pathological circumstances [12].

More specifically, a decreased activation response of thrombocytes may result in excessive bleeding, whereas platelet hyperactivity may lead to thrombotic complications. Additionally, platelet hyperactivity may be associated with various processes such as cancer metastasis, angiogenesis, the alteration of liver tissue, atherosclerosis, and an inflammatory and immune response [13–16], as well as many other platelet-mediated diseases, namely, cardiovascular disease (CVD) [17–19], Alzheimer's disease, renal diseases, arterial and venous thrombosis, and microorganism infections [14,16,20–22]. However, the explanatory mechanisms have not been well studied. Thus, the assurance of regular thrombocyte aggregation is essential in averting platelet-related diseases in the conventional human population [23–27].

Against this background, the exception should also be made to the athletes' population, where the activity of the bodily endothelial cells in athletes is rigorously related to the function of thrombocytes. In an abnormal state, the endothelial dysfunction can magnify the endothelial synergy with platelets [28]. Therefore, regular physical activity promotes blood circulation and shear stress, resulting in the increased bioavailability of nitric oxide (NO), which, in turn, is accompanied by its beneficial effects on endothelial function [29]. As a consequence, the occurrence of more intense blood flow and fluctuations in hemodynamics triggered by high physical loads may lead to both chronic and acute alterations in vascular activity.

High-intensity training, especially in some sports, can give rise to immoderate platelet aggregation and increase the risk of blood clot events. Science has long considered marathons and triathlons as sports related to a certain risk for thrombotic events [30]. Acute strength exercises may contribute to platelet activation, too [31]. Although the promotion of endothelial function has been demonstrated by an increased NO availability, the exact pathways underlying this phenomenon remain unresolved. However, these exercise-related outcomes associated with the platelet function are clearly similar to a well-established relationship between workouts and the risk for adverse cardiovascular events, where strenuous physical loads temporarily increase the risk for cardiac infarction while a healthy lifestyle dramatically diminishes deaths from heart and circulatory disorders [31].

As a result of the above-mentioned, there is a sudden and noticeable increase in focus on naturally found antiplatelet inhibitors that humans can use habitually. Research has identified a large number of dietary antiplatelet nutrients that may decrease the hyperactivity of platelets without any adverse effects [32–36]. For example, in terms of human nutrition, dietary fatty acids (FAs) can either trigger or inhibit platelet aggregation. Omega-3 FAs, mainly found in fish oil, has been shown to wield antiplatelet effects. Dietary omega-3 long-chain polyunsaturated FAs regulate platelet activity through the thromboxane B3 (TxB3) synthesis, the alteration of platelet membranes, and the acceleration

Nutrients 2025, 17, 3418 3 of 17

of NO synthesis [14,32,37]. Therefore, these useful omega-3 FAs incorporate into platelet cell membranes, modifying their functionality and fluidity [38]. On the other hand, proinflammatory omega-6 FA, mainly found in vegetable oils as well as in seeds, eggs, nuts, poultry, and whole grains, can stimulate platelet aggregation.

Generally, it is important to highlight that the positive adaptive response, along with phenotypic alterations, depends upon increased performance, which in turn relies on the appropriate recovery time between each training course, too. High-performance athletes receive annual training and participate in competitions with periods of recovery that are not always suitably adapted to the workload. Thus, in view of the interrelation between the training courses, the nutritional status, and the variations in essential blood parameters, it is important to detect all the pathways in which the adaptation of the athlete's body to physical loads can be optimized. In this connection, although the actual dietary habits play a vital role in determining the platelet function, a core element in the body's clotting process [1], the outstanding relationship between the platelet function and the macronutrient intake amongst high-performance athletes remains largely unanalyzed and unexplained at present. For sequential arrangements, to fill the existing specific scientific gaps, this study aimed to explore the association between dietary fat intake and the indices of blood profiles, concentrating on platelet variables in a sample of high-performance athletes.

# 2. Materials and Methods

# 2.1. Study Design, Population, and Data Collection

A single cross-sectional study in design was conducted at the Lithuanian Sports Centre (LSC) between April and August in 2021–2022. A representative sample size (n = 75–176) with a constructed confidence interval of a 95% confidence level and a marginal error of 5–10% was obtained from the eligible population (N = 322) of elite athletes when applying the web-based online freeware OpenEpi version 3.01 [39].

Out of the eligible population, the following 176 high-performance athletes were included in the study using a probability sampling method, taking into account the estimated representative sample size and the inclusion criteria as follows: (1) athletes who trained for the Olympics; (2) study participants who represented a country as national or international athletes; (3) athletes who engaged in a high-frequency, six-day-per-week training course; (4) athletes in the preparation phase.

The athletes were collected from the authorized list established by the National Olympic Committee of Lithuania (LTOK), the authorized body responsible for the Olympic movement in Lithuania [40]. The exclusion criteria were personalized as follows: (1) athletes who were vigorously competing in their respective sports during the study period (n = 19); (2) athletes who have reported muscle, tendon, or ligament injuries (e.g., hamstring strains, Achilles tendon issues, ankle sprains) (n = 5); (3) female athletes throughout their regular menstrual cycle (n = 12); (4) specific athletes who abstained from participating in the cross-sectional study (n = 58). Hence, healthy and non-medicated athletes were included in the study. A more detailed scheme for the enrollment process is displayed in Figure 1. Finally, based on a priori metabolic pathways (in terms of aerobic and anaerobic) in the body [41], the professional athletes (n = 82) were divided into two groups as follows: aerobic athletes ((n = 48); swimmers (n = 13), rowers (n = 2), modern pentathlon players (n = 5), road cyclists (n = 7), skiers (n = 8), biathletes (n = 7) and boxers (n = 6)) and anaerobic athletes (n = 34): Greco-Roman wrestlers (n = 8), judokas (n = 8), and basketball players (n = 18)).

Nutrients 2025, 17, 3418 4 of 17

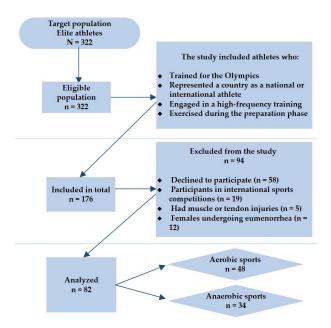


Figure 1. Enrollment flowchart for high-performance athletes.

### 2.2. Measurements

# 2.2.1. Dietary and Body Weight Assessment

The assessment of the athletes' nutritional profile was performed using a 3-day food record approach where high-performance athletes tracked all food, dishes, and beverages over three consecutive days, aiming to evaluate a typical dietary intake and the levels of macronutrients consumed [42]. The specific food items, including portion sizes (using weight or measures), cooking approaches, food brands, and meal timing, consumed by athletes were recorded by a sports nutritionist through direct interviews [43], while the special Atlas of Foodstuffs and Dishes [44] for research purposes was also used. The energy intake (EI) and dietary macronutrient (carbohydrates, protein, lipids, saturated fatty acids (FA), cholesterol, monounsaturated FA, polyunsaturated FA, omega-6 FA, and omega-3 FA) intakes were assessed via the Nutrition Baseline software (NutriSurvey, the English translation of a commercial German software (EBISpro version 3.02), SEAMEO-TROPMED RCCN-University of Indonesia) [45]. The typical food dataset of NutriSurvey was manually augmented by the nutritional values of foodstuffs extracted from the Lithuanian chemical composition tables [46].

The intakes of macronutrients by the high-performance athletes we studied were evaluated in agreement with the Recommended Dietary Allowances (RDAs) established by the scientific recommendations [47–51]. Therefore, the recommended carbohydrate intake for athletes must be equal to 7–10 g/kg of body weight (BW), and protein intake must comprise 1.2–2.0 g/kg of BW. The total dietary fat percentage of EI should fluctuate from 25 to 35%, the saturated FA percentage must be <10% of EI, the polyunsaturated FA percentage should range from 6 to 10% of EI, the omega-6 FA percentage must vary between 5 and 8% of EI, omega-3 FA percentage should be  $\geq$ 2% of EI, and the omega-6 FA to omega-3 FA ratio should be in a healthy range considered to be between 4:1 and 1:1 [52].

For comparison, our study rated the EI of athletes with the daily energy expenditure, basal metabolic rate, energy expenditure during training exercises, and the daily energy expenditure of all study participants. The basal metabolic rate was calculated applying the Harris and Benedict equations [53]. The levels of physical activity were assessed according to the recommendations along with the evaluation algorithm established by the American College of Sports Medicine and the American Dietetic Association, Dietitians of Canada [54]. The benchmark data of the basic physical exertion and the activity codes, coupled with the

Nutrients 2025, 17, 3418 5 of 17

metabolic equivalents (in kcal/kg/h) for specific physical activities, were applied in the observational study. These proceedings were supplemented by the research of Ainsworth et al. [55], and the data were handled depending on the specific activity. Thus, the physical activity level was calculated as the ratio between the daily energy expenditure and the basal metabolic rate, as mentioned by FAO/WHO/UNU [56]. Eventually, in order to calculate the total energy requirement, the basal metabolic rate was then multiplied by the suitable physical activity factor.

The measurements of BW (in kg) and standing height (achieving  $\pm 1$  cm accuracy) were assessed using the bioelectrical impedance analysis (BIA) via the body composition analyzer X-Scan Plus (the International Organization for Standardization adopted by the European Union (EN-ISO): 13488 [57], Seoul, Republic of Korea).

## 2.2.2. Blood Collection

During the periodical medical examination, blood samples were collected from the selected athletes by a trained phlebotomist at an accredited LSC clinical laboratory (medical laboratory license number 2545; Vilnius, Lithuania). In order to control the possible effect of exercise on the hematological results, study participants were asked to avoid physical effort for 24 h before phlebotomy, as well as to be hydrated and fasted.

In laboratory settings, the hematology profile of athletes (red blood cell (RBC) count), red blood cell distribution width (RDW), hemoglobin (Hb), mean corpuscular hemoglobin concentration (MCH), hematocrit (Ht), white blood cell (WBC) count, lymphocytes (Lym), monocytes (Mo), platelet (PTL) count, mean platelet volume (MPV), and plateletcrit (PCT)) was assessed via the Nihon Khoden automated hematology analyzer (MEK6400-K, Tokyo, Japan). Also, for the study data analysis, an additional platelet function-related variable, namely, the platelet-to-hemoglobin ratio (PHR), was calculated by dividing the platelet count by the hemoglobin concentration [58,59].

The blood samples of study participants were tested individually, with the possibility of repeating and verifying the unusual results. The laboratory monitored the precision and accuracy of all blood samples analyzed.

## 2.3. Statistical Data Analysis

The statistical examination was completed utilizing the Statistical Package for the Social Sciences (IBM SPSS Statistics) version 25.0 for Windows (IBM Corp, Armonk, NY, USA).

Firstly, the Kolmogorov–Smirnov test as an a priori statistical approach was used to test for data normality. In the second place, the primary empirical data obtained from this study were classified according to both bivariate and multivariate analyses.

For the bivariate analysis, the means  $\pm$  standard deviations (SDs) for normally distributed data were calculated, and a t-test, along with Cohen's D (d) effect sizes, including the Pearson's correlation coefficient (r), was used for data analysis. More specifically, nutritional adequacy was verified by contrasting macronutrient intakes to the RDAs. A t-test coupled with d estimates [60] ('trivial effect':  $0 \le d < 0.2$ ; 'small effect':  $0.2 \le d < 0.5$ ; 'moderate effect':  $0.5 \le d < 0.8$ ; 'large effect':  $0.8 \le d$ ) as effect sizes were applied to resolve whether there was a significant difference between the means of the two subgroups (e.g., EI and the essential macronutrient intake amounts vs. the daily energy expenditure and RDAs for macronutrients). Also, the t-test was used to determine whether the athletes competing in different sports (aerobic vs. anaerobic) had different nutritional and hematological profiles. In addition, in order to assess the correlation between the PTL count ( $\times 10^9/L$ ) and the MPV (fL) and Hb (g/dL), the r values were calculated and interpreted as follows: 'negligible correlation' ( $0 \le r < 0.3$ ); 'low to moderate positive correlation' ( $0.3 \le r < 0.7$ ); 'high to very high positive correlation' ( $0.7 \le r \ge 0.9$ ).

Nutrients **2025**, 17, 3418 6 of 17

For multivariate analysis, our study carried out multiple logistic regression analyses with the binary dependent variables around four groups covering the indices of blood profiles, namely, '244  $\times$  10 $^9/L$   $\leq$  PTL count > 244  $\times$  10 $^9/L'$ , '0.19%  $\leq$  PCT > 0.19%', '14.7 g/dL  $\leq$  Hb > 14.7 g/dL', '16.4  $\leq$  PHR > 16.4'. The independent variables, namely, '3307 kcal/day  $\leq$  EI > 3307 kcal/day', '395.4 g/day  $\leq$  carbohydrates > 395.4 g/day', '115.6 g/day  $\leq$  protein > 115.6 g/day', '143.9 g/day  $\leq$  total fat > 143.9 g/day', '50.3 g/day  $\leq$  saturated FA > 50.3 g/day', '720 mg/day  $\leq$  cholesterol > 720 mg/day', '1.04 g/day  $\leq$  omega-3 FA > 1.04 g/day', '19.9 g/day  $\leq$  omega-6 FA > 19.9 g/day' were adjusted to the dichotomous form. Thus, the units of measurement of dependent and independent variables were modified to the dichotomous form based on cut-off values that were set by referring to the median values of the group of athletes studied. All logistic regression models were adjusted for the sex and sports discipline of professional athletes.

In all the statistical tests applied to data analysis, a two-tailed p-value  $\leq 0.05$  was considered statistically significant. The SPSS software, along with the free and open-source software LibreOffice version 7.6.4, was used for the visualization of statistical data.

# 3. Results

#### 3.1. Characteristics of Athletes

Out of 82 high-performance athletes of a mean age of  $19.8\pm2.2$  years who constituted the representative sample, 25.6% were females and 74.4% were males. From a sporting perspective, study participants were divided into aerobic and anaerobic athletes at 58.5% and 41.5%, respectively. The range for training practice equaled  $6.8\pm2.5$  years, the number of training sessions was 6 days per week, and the duration of daily workout was  $173.2\pm66.3$  min on average. The mean standing height and body weight of the athletes involved in the study were  $181\pm13$  cm and  $71.6\pm15.4$  kg, respectively. The extent of the workload of the athletes was in agreement with the validated training plans. For the purpose of assessing the workload plans of the athletes studied, account has been taken of the training plans for athletes that were officially confirmed by the LTOK. More detailed data referring to the exercise plans designed for high-performance athletes is provided in Table 1.

**Table 1.** The exercise plans for high-performance athletes.

Sports	Skiing and Biathlon	Boxing, Graeco-Roman Wrestling and Judo	Road Cycling	Swimming (50–400 m) and Canoe Paddling (500–2000 m)	Modern Pentathlon	Basketball
Performance investigation by month	August	May–June	April	March-May	June	May
Macrocycle phase Stage Days of exercise per month	Preparation Basic training 27	Preparation Special training 27	Preparation Special training 26	Preparation Special training 25–27	Preparation Special training 26	Preparation Special training 27
Total physical training (hours per month)	31–32	36-41	36	40–41	40	41
Special training is categorized into five zones of intensity depending on ATP production in the muscles						
Zone 1: Aerobic strength endurance training, recovery (pulse rate $-130 \pm 10$ bpm, lactate level up to 2 mmol/L)	26–36%	12–18%	18%	18–20%	20%	18%
Zone 2: Aerobic strength training (pulse rate $-150\pm10$ bpm, lactate level is 2–4 mmol/L), muscular power increase at the anaerobic threshold	37–48%	26–30%	50%	40-50%	50%	42%
Zone 3: Mixed aerobic and anaerobic glycolytic strength training (pulse rate $-170\pm10$ bpm, lactate level is $4$ – $12$ mmol/L), increase of VO <sub>2</sub> max	15–17%	40–50%	32%	29–35%	25%	32%

Nutrients **2025**, 17, 3418 7 of 17

Table 1. Cont.

Sports	Skiing and Biathlon	Boxing, Graeco-Roman Wrestling and Judo	Road Cycling	Swimming (50–400 m) and Canoe Paddling (500–2000 m)	Modern Pentathlon	Basketball
Zone 4: Anaerobic glycolytic strength training (pulse rate ≥181 bpm, lactate level up to 21 mmol/L)	6–10%	8–10%	-	5%	3%	5%
Zone 5: Anaerobic phosphocreatine strength training (lactate level is 1.5–6 mmol/L)	2–3%	2–4%	_	1–2%	2%	3%

The exercise time was assigned for intensity zones (%); ATP—adenosine triphosphate.

#### 3.2. Nutritional Status

Table 2 presents the reported usual mean intakes of energy, carbohydrates, protein, fats, saturated FAs, cholesterol, monounsaturated FAs, and polyunsaturated FAs in high-performance athletes.

**Table 2.** Macronutrient intakes in high-performance athletes according to sport branches.

Energy Intake and Macronutrients	Endurance Athletes (n = 48)	Strength-Power Athletes $(n = 34)$	RDAs	đ
Energy intake (kcal/kg/day)	$49 \pm 15$	$50 \pm 16$	$50 \pm 8  [49; 53]$	-0.07
Carbohydrates (g/kg/day)	$5.5 \pm 1.9$	$5.3 \pm 1.9$	7–10	-1.63
Carbohydrates (% of EI)	$47.3\pm7.4$ **	$43.2 \pm 6.2$	45–65	-1.31
Protein (g/kg/day)	$1.6 \pm 0.5$	$1.7\pm0.5$	1.2-2.0	-0.02
Protein (% of EI)	$14.2 \pm 3.1$	$14.0 \pm 2.6$	12-20	-0.82
Lipids (g/kg/day)	$2.0 \pm 0.8$	$2.3 \pm 0.8$	1.0 - 1.5	1.06
Lipids (% of EI)	$38.5 \pm 6.9$	42.8 $\pm$ 5.7 **	25–35	1.53
Saturated FA (g/day)	$45.5\pm19.7$	$60.4\pm20.5$ ***	_	_
Saturated FA (% of EI)	$13.4 \pm 3.4$	$14.3 \pm 2.5$	≤10	1.22
Cholesterol (mg)	$698 \pm 391$	982 $\pm$ 478 ***	_	_
Monounsaturated FA (g/day)	$63.5 \pm 23.6$	97.2 $\pm$ 47.6 ***	_	_
Polyunsaturated FA (g/day)	$20.1 \pm 10.7$	27.2 $\pm$ 12.0 **	_	_
Polyunsaturated FA (% of EI)	$5.9 \pm 2.4$	$6.3\pm1.7$	6–10	-1.86
Omega-6 FA (g/day)	$18.4 \pm 9.8$	$25.4 \pm 11.5$ **	_	_
Omega-6 FA (% of EI)	$5.4\pm2.2$	$5.9\pm1.7$	5–8	-0.19
Omega-3 FA (g/day)	$1.1\pm0.8$	$1.3 \pm 0.5$	_	_
Omega-3 FA (% of EI)	$0.3 \pm 0.2$	$0.3 \pm 0.1$	$\geq$ 2	-8.50
Omega-6 FA/Omega-3 FA ratio	$18.9 \pm 8.1$	$19.4 \pm 6.1$	1:1-4:1	2.07

Data are presented as means  $\pm$  standard deviations (SDs). The measure of Cohen's D displays the effect size (d) of the mean differences in dietary nutrient intakes after equating them to the RDAs. FA—fatty acids, RDA—recommended dietary allowance. All p-values (p) obtained from the t-test reporting the significance of differences between the dietary nutrient intakes depending on the sport branches. \*\*— $p \le 0.01$ , \*\*\*— $p \le 0.001$ .

For elite athletes, the recorded mean consumption levels of energy, carbohydrates, protein, and fat were  $49 \pm 15$  kcal/kg/day,  $5.4 \pm 1.9$  g/kg/day,  $1.6 \pm 0.5$  g/kg/day, and  $40.3 \pm 6.7\%$  of energy intake, respectively. Overall, the comparison of the results with RDAs showed that the levels of the basic macronutrients consumed by athletes were not balanced, i.e., carbohydrate intake was too low (d = -1.63) while fat consumption was excessive (d = 1.06). For anaerobic athletes, the intakes of saturated FA and monounsaturated FA and dietary cholesterol were expressed at higher levels compared to those in the athletes competing in endurance sports ( $0.001 \le p \le 0.05$ ). Specifically, in aerobic and anaerobic athletes, the mean daily cholesterol intake ranged from  $698 \pm 391$  mg to  $982 \pm 478$  mg, respectively. The dietary cholesterol intake of this magnitude, usually originating from a typical meal high in saturated fat, exceeded the previously existing RDA [61] for cholesterol (300 mg/day) by 2–3 times.

Nutrients 2025, 17, 3418 8 of 17

On the contrary, in terms of effect sizes (d), a severe deficiency of polyunsaturated FA and omega-3 FA was observed in athletes' nutrition (d = -1.86 and d = -8.50). In addition, when the optimum ratio of omega-6 FA and omega-3 FA should be 4:1, this study identified an excess ratio (19.1:1) of almost five times the recommended one, regardless of the sport being cultivated by athletes.

# 3.3. Blood Analysis

Table 3 shows the analysis of the blood samples of both aerobic and anaerobic athletes studied. Essential hemogram parameters (RBC count, Hb, MCV, Ht, MCH, WBC count, Lym, Mo, PTL count, PCT, MPV) in athletes were within the normal range. Taking into account the variations found in the trivial effect size (d) from -0.2 to 0.19, it can be confirmed that the baseline blood parameters fluctuated within the standards, especially when compared to the recommended lower bound (2.5th percentile) and upper bound (97.5th percentile) of the RIs and their 90% CIs established for the athletes' hematological parameters [62]. On the other hand, the analysis of blood samples revealed that red cell distribution width (RDW  $11.6 \pm 0.7\%$ ) was below the minimum recommended threshold (RDW 12.1-14.3) for athletes (d=-2.3). Thus, when this score exceeded the recommended range, there was a possibility that study participants had a nutrient deficiency.

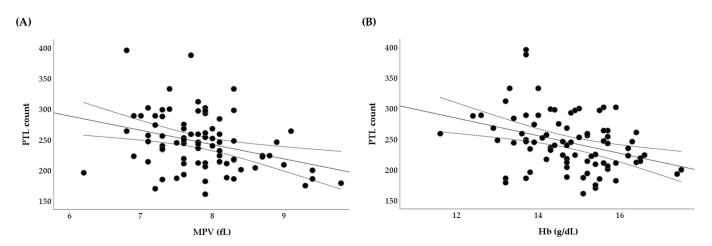
Table 3. Hemogram of elite athletes.

Parameter	Endurance Athletes (n = 48)	Strength-Power Athletes $(n = 34)$	RIs for Physically Active Individuals 2.5th–97.5th Percentile	d
RBC count ( $\times 10^{12}/L$ )	$5.1 \pm 0.4$	$5.0 \pm 0.5$	4.4–5.6	0.14
Hb (g/dL)	$14.7\pm1.1$	$14.9\pm1.2$	13.0–16.1	0.17
MCV (fL)	$86.4 \pm 3.4$	$87.2 \pm 3.8$	80.9–94.9	-0.19
Ht (%)	$44.4 \pm 3.7$	$44.0 \pm 3.7$	39.5–48	0.13
MCH (pg)	$28.6 \pm 1.5$	$29.4 \pm 1.6$	26.1–31.6	0.01
RDW (%)	$11.7\pm0.8$	$11.6 \pm 0.7$	12.1–14.3	-2.3
WBC count ( $\times 10^9/L$ )	$6.9 \pm 1.6$	$7.2 \pm 1.9$	4.5–10.1	-0.17
Lym (%)	$34.0 \pm 6.3$	$34.5 \pm 8.4$	20–44	0.19
Mo (%)	$6.9 \pm 1.8$	$6.6 \pm 1.6$	2.0-9.5	0.11
PTL count ( $\times 10^9/L$ )	$247\pm49$	$241 \pm 42$	140–337	0.10
PCT (%)	$0.19 \pm 0.03$	$0.20 \pm 0.03$	0.12-0.41	-0.20
MPV (fL)	$7.8 \pm 0.6$	$7.9 \pm 0.7$	5.9–9.9	0.14

Data are presented as means  $\pm$  standard deviations (SDs). Hb—hemoglobin; Ht—hematocrit; Lym—lymphocytes; MCH—the mean corpuscular hemoglobin concentration; Mo—monocytes; MPV—the mean platelet volume; PCT—plateletcrit; PTL—platelet; RBC—red blood cells; RIs—reference intervals [62]; RDW—the red blood cell distribution width; WBC—white blood cells; d—the effect size.

The results of this study showed a reverse correlation between the PTL count and the MPV (r = -0.3, p = 0.003) (Figure 2A). It should be highlighted that the mean platelet volume (MPV) is a measurement of the mean platelet size. Although it is an indicator of platelet activation, a significant inverse correlation between the MPV and PTL count can be found [63]. In healthy athletes and under certain conditions, lower platelet counts may be related to higher MPV, and vice versa, taking into account the fact that the medulla ossium responds to a greater demand by releasing younger and larger thrombocytes. In other words, lower levels of platelet count and higher levels of MPV may serve as signs of the athletes moving to a better recovery and regeneration between the intense physical loads [64].

Nutrients 2025, 17, 3418 9 of 17



**Figure 2.** (A,B) The correlation between the platelet (PTL) count ( $\times 10^9$ /L) and the mean platelet volume (MPV) (fL), and hemoglobin (Hb) (g/dL).

In addition, our study explored the platelet-to-hemoglobin ratio (PHR =  $16.8 \pm 3.9$ ) as a novel indicator for stress and inflammation [58,59]. In this case, higher PHR may imply increased PTL counts and/or decreased Hb levels, to which the athlete's body may respond with increased inflammation induced by physical exertion or chronic stress. Thus, the results obtained from our study have confirmed the inverse correlation between PTL and Hb (r = -0.4, p = 0.001) (Figure 2B). This study, in its subsequent stages, assessed a possible relationship between the nutritional status and the indices of the athletes' blood profiles, including the platelet-to-hemoglobin ratio.

# 3.4. Associations Between Dietary Intakes and Platelet Variables

As shown in Figure 3A–D, further multivariate logistic regression analyses revealed significant relationships between the dietary intakes of polyunsaturated FA and the indices of blood profiles in a cohort of elite athletes. Following the adjustment of logistic regression models for athletes' sex and sports discipline [65–67], in relation to slightly higher levels of blood PTL counts (>244  $\times$  10<sup>9</sup>/L), the AOR for higher intake of omega-6 FA among high-performance athletes was 6.8 (95% CI 1.8; 10.2, p = 0.016). Similarly, an association was also found between PCT (>0.19%) and omega-6 FA intakes (AOR 9.5, 95% CI 1.2; 9.9, p = 0.029).

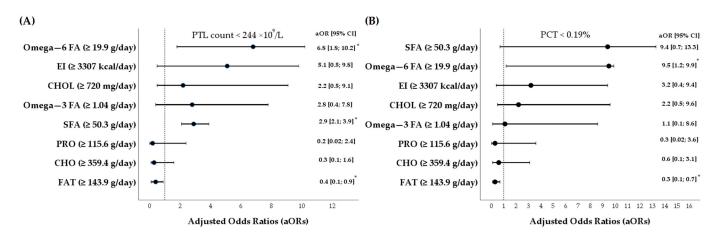
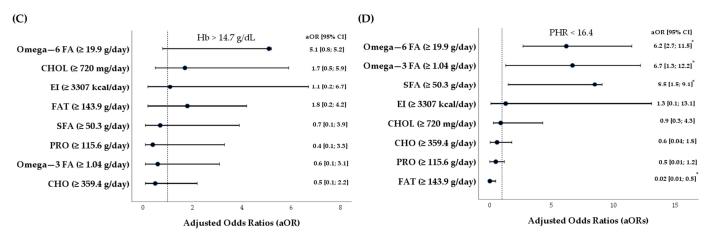


Figure 3. Cont.

Nutrients 2025, 17, 3418 10 of 17



**Figure 3.** (A–D) A graphic representation of multivariate logistic regression models (dependent variables: PTL count ( $\times 10^9$ /L), PCT (%), Hb (g/dL), and PHR; independent variables: EI (kcal/day), CHO (g/day), PRO (g/day), FAT (g/day), SFA (g/day), CHOL (mg/day), omega-6 FA (g/day), and omega-3 FA (g/day). A dotted line represents the adjusted odds ratio (AOR) equal to 1. If AOR < 1 and AOR  $\neq$  1, it means that there is an inverse association between the independent and dependent variables. If AOR > 1 and AOR  $\neq$  1, it means that there is a positive association between the independent and dependent variables. The multivariate logistic regression models (A–D) were adjusted for athletes' sex and sports discipline. \*— $p \le 0.05$ , 95% CI—95% confidence interval, CHO—carbohydrates, CHOL—cholesterol, EI—energy intake, FAT—fats, PCT—plateletcrit; PHR—the platelet-to-hemoglobin ratio; PTL—platelet, PRO—protein, FA—fatty acids, SFA—saturated FA.

Finally, the PHR (<16.4), as a blood-based biomarker calculated by dividing PTL counts by Hb levels and used for an indirect assessment of systemic stress, immune activity, and inflammation [58,59], was inversely associated with higher intake levels of dietary omega-3 FA (AOR 6.7, 95% CI 1.3; 12.2, p = 0.029), omega-6 FA (AOR 6.2, 95% CI 2.7; 11.5, p = 0.009), and saturated FA (AOR 8.5, 95% CI 1.5; 9.1, p = 0.020) (Figure 3D).

#### 4. Discussion

The aim of this study was to assess the levels of macronutrients consumed by high-performance athletes and to associate the latter with the basic indicators of a hemogram. The study found that macronutrients, namely, protein and carbohydrates, consumed by athletes were not associated with the platelet count, plateletcrit, or platelet-to-hemoglobin ratio. Nevertheless, within the segment of elite athletes under analysis, our study observed the imbalances between the intake of basic macronutrients, manifested by the deficit consumption of carbohydrates (5.3–5.7 g/kg of body weight) and an excessive consumption of the total dietary fat (38.5–42.8% of EI), exclusively saturated FA (13.4–14.3% of EI), and dietary cholesterol (698–982 mg/day). A typical isocaloric Western-type diet consumed by athletes contained a higher percentage of calories from omega-6 FA (around 5.4–5.9% of EI) and a lower percentage from omega-3 FA (around 0.3% of EI). Given that the ideal human omega-6 to omega-3 FA ratio is commonly deemed to be between 1:1 and 4:1, an athlete's 'Western diet' was heavily skewed, with a ratio fluctuating from 18.9:1 to 19:4 in favor of omega-6 FA.

Our study revealed a positive association between the omega-3 FA intake and the platelet-to-hemoglobin ratio, which has been indirectly related to the athletes' poor nutrition status, inflammation, and oxidative stress. On the other hand, although this study did not reveal a positive relationship between the omega-3 FA consumption and the total platelet count or plateletcrit, our findings were partly consistent with the data collected from the scientific research, with scientists speculating that FA obtained from a healthy or balanced diet can trigger or inhibit platelet aggregation. Thus, omega-3 FA, mainly

Nutrients 2025, 17, 3418 11 of 17

found in fish oil and walnuts, such as eicosatetraenoic acid (EPA) and docosahexaenoic acid (DHA), have been demonstrated to boost the antiplatelet function and the immunity [68–73]. More specifically, these useful FAs can integrate into the membranes of platelet cells and subsequently modulate their fluidity and activation [38]. This antiplatelet effect of omega-3 FA can be explained by the fact that EPA and DHA can change the membrane fluidity, the expression of a regulating receptor, and the production of thrombin [74]. Resolvins (resolvins E and D), as endogenous lipid mediators derived from omega-3 FAs, are capable of decreasing and inhibiting thromboxane-induced thrombocyte aggregation. Although the antiplatelet effect of oils or marine fish has been confirmed in healthy study participants, the dose used was considered to be a determining factor in serving a benefit, especially given that the low and high doses do not always indicate a benefit [37,75–82].

Also, research-based evidence has revealed that the increase in the omega-6 to omega-3 FA ratio can shift the balance into a proaggregatory state [83]. This study revealed similar findings in relation to the excessive ratio of omega-6 to omega-3 FA resulting from higher consumption of omega-6 FA among high-performance athletes. In addition, when higher platelet counts are linked to an increased risk of clotting, our study found an association between the higher intake of omega-6 FA and the total platelet count as well as plateletcrit. Thus, higher consumption of omega-6 FA may serve as the potential precursor in blood coagulation and vasoconstriction. In this context, unconstrained or disproportionate blood clotting is not appropriate for athletes, as it may elevate the risk of serious blood clots (thromboembolism) depending on conditions, namely, muscle damage, dehydration, psychological stress, or immobility during travel, regardless of the healthy nature of athletes.

Furthermore, this study showed that lower levels of the platelet-to-hemoglobin ratio were related to an increased consumption of omega-3 FA, omega-6 FA, and saturated FA in high-performance athletes under our analysis. Preliminary, these findings refer to the possible fact that the activity of the immune system is not impaired by the chemical nature of the FA consumed by elite athletes. In agreement with these findings, it should be noted that as the predecessor to pro-inflammatory eicosanoids, high levels of omega-6 FA intake cannot directly cause inflammation, mostly in healthy people; however, high omega-6 FA intake itself may restrict the anti-inflammatory outcomes from the omega-3 FA. Thus, in the context of inflammation, the relationship between omega-3 FA and omega-6 FA and their lipid intermediaries remains mixed and still misunderstood, especially in athlete populations.

Additionally, the results originating from our study did not match the authors' reports that food rations high in saturated fats may amplify the platelet function, bringing about the pro-thrombotic conditions [84]. These contradictory results can be explained by the disparities observed in populations of non-sporting people and professional athletes. More specifically, in terms of the non-physically active population, a high-fat diet may increase the levels of 'bad' low-density lipoprotein (LDL) cholesterol, resulting in hyperlipidemia, which, in turn, can promote platelet activity. Secondly, the increased platelet activation may take place via the occurrence of changes in platelet phospholipid content and the activation of signaling pathways similar to P2Y12 [85,86]. In this case, it should be highlighted that almost all of the athletes involved in our study reported high consumption levels of fat and saturated fatty acids. Other researchers have revealed the same findings [87]. However, to date, there has been no scientific evidence that a high-fat diet leads to overweight athletes [88]. On the contrary, there is evidence that the use of a high-fat diet can increase the levels of circulating homocysteine, which serves as a potential risk factor for the development of cardiometabolic disorders [89]. The athletes' demand to reduce the consumption of saturated FA may also be confirmed by the results obtained from other empirical studies, Nutrients 2025, 17, 3418 12 of 17

which refer to the fact that high intakes of saturated FA in athletes were related to greater muscle soreness and impaired recovery by triggering adipose tissue inflammation via receptors like TLR4 [90].

Therefore, the athletes may reduce this potential inflammation by using polyunsaturated FA such as omega-3 FA. Also, reducing the consumption of foods rich in saturated FA and increasing the consumption of whole grains, low-fat dairy products, lean poultry and meats, fish, fruits, and vegetables could bring more nutritional benefits to athletes in order to enhance the adaptation to intense physical loads [28,90]. Within this framework, the cohorts of athletes become distinctive in line with the International Society of Sports Nutrition (ISSN) position stand, establishing the following: (1) 'Athletes may be at a higher risk for omega-3 FA insufficiency'; (2) 'Diets rich in omega-3 FA, including supplements, are effective strategies for increasing omega-3 FA levels'; (3) 'Also, omega-3 FA supplementation can positively affect various immune cell responses in athletic populations' [73].

This study had several limitations. In particular, minor biases could have occurred due to the methodology used in the dietary assessment process, on the basis of which the accuracy of the final dataset could have been affected by specific factors such as the training status, nutrition-related beliefs, the level of competition, or memory gaps in the composition of the diet consumed or recognizing food brands. Secondly, the search for associations between independent variables and platelet variables may have been partially distorted by potential confounders related to protein consumption. More precisely, some research has found that dietary proteins, particularly those rich in certain amino acids like glycine and arginine, and platelet function were interconnected [71,72]. Thirdly, the relatively moderate, although representative, sample size of our study can restrict the extrapolation of the results of this study to a less physically active segment of the human population. Fourthly, given that this exploratory study used a cross-sectional analysis, the results obtained from this study cannot refer to causal relationships between the variables analyzed. In this case, taking into account our novel study findings, undertaking further longitudinal and/or experimental studies in design could provide additional pronounced scientific robustness. Also, the additional measurement of biomarkers (i.e., the omega-3 index on erythrocytes and lipid profiles on the plasma) could make the correlations with the hematological indicators more precise.

# 5. Conclusions

Considering that increased physical loads, even within the limits of physiological norms, may have an adverse impact on both the total platelet count and their functional activity, the results of this study highlighted important associations between the dietary fat profile and the platelet-related variables observed in a sample of high-performance athletes.

Following the adjustment to athletes' sex and sports discipline, the outcomes related to slightly higher levels of blood platelet counts and plateletcrit were associated with higher intakes of omega-6 fatty acids. Moreover, a higher platelet-to-hemoglobin ratio, as a novel indirect blood-based biomarker pronouncing potential inflammatory processes in the body, had a reverse relationship with higher intake levels of dietary omega-3, omega-6, and saturated fatty acids among elite athletes.

In this context, whilst high-performance athletes consume high levels of saturated fatty acids, including dietary cholesterol, and low omega-3 fatty acids during their training process, this dietary behavior related to the intake of different dietary fatty acid fractions still maintains unrecognized metabolic pathways linked to the possible positive benefits or negative outcomes for the immune system, especially in the event of progressive high physical workloads among elite athletes. Thus, the findings originating from this study may benefit the athletes exposed to extreme physical exposure, coaches, and healthcare

Nutrients 2025, 17, 3418 13 of 17

providers, or initiate further research into health-related consequences of the intakes of both polyunsaturated fatty acids and saturated fatty acids, including dietary cholesterol.

Finally, a holistic understanding of the platelet function, which exclusively includes the role of nutrition, can enhance the perception of thrombosis and shed light on the development of new intervention or treatment strategies. The prospect of personalized nutrition targeted at the professional athletes' segment may provide an innovative opportunity to increase their capacity to manage platelet function via diet, while stressing the importance of further empirical experimental research in this dynamic and vital biomedical field, too.

**Author Contributions:** Conceptualization, M.B., R.S. and I.K.; methodology, M.B., I.K. and R.S.; software, M.B. and J.L.; validation, M.B., I.K., J.L. and R.S.; formal analysis, M.B. and R.S.; investigation, M.B. and R.S.; resources, J.L. and I.K.; data curation, I.K. and R.S.; writing—original draft preparation, M.B., I.K., J.L. and R.S.; writing—review and editing, M.B., I.K., J.L. and R.S.; visualization, M.B. and J.L.; supervision, J.L., I.K. and R.S.; project administration, I.K. and J.L. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

**Institutional Review Board Statement:** This study was conducted in accordance with the Declaration of Helsinki and approved by both the Institutional Review Board of the State Higher Education Institution Panevėžys College (protocol code BMF2-10, protocol approved on 10 March 2021) and the Vilnius Regional Committee of Biomedical Research Ethics (five-year protocol code 158200-17-898-419, of 11 April 2017).

Informed Consent Statement: Informed consent was obtained from all subjects involved in the study.

**Data Availability Statement:** The data presented in this study are available on request from the corresponding author due to sharing them would go against ethical considerations, such as study participants' privacy and consent.

**Conflicts of Interest:** The authors declare no conflicts of interest.

## **Abbreviations**

ATP	Adenosine triphosphate
BIA	Bioelectrical impedance analysis
BW	Body weight
CI	Confidence interval

CVD Cardiovascular disease
DHA Docosahexaenoic acid
FI Energy intake

EI Energy intake
EN European Union
EPA Eicosapentaenoic acid

FA Fatty acids
Hb Hemoglobin
Ht Hematocrit

ISO International Organization for Standardization
ISSN International Society of Sports Nutrition

LB Lower bound

LDL Low-density lipoprotein

Lym Lymphocytes

LSC Lithuanian Sports Centre (LSC)

LTOK Lithuanian National Olympic Committee
MCH Mean corpuscular hemoglobin concentration

Nutrients 2025, 17, 3418 14 of 17

Mo Monocytes

MPV Mean platelet volume

NO Nitric oxide
OR Odds ratio
PCT Plateletcrit

PHR Platelet-to-hemoglobin ratio

PTL Platelet

RBC Red blood cells

RDA Recommended dietary allowance RDW Red blood cell distribution width

SD Standard deviation

SPSS Statistical Package for the Social Sciences

TxB3 Thromboxane B3
UB Upper bound
WBC White blood cells

## References

1. Roy, S.; Paul, K. Nourishing the clot: A comprehensive review of dietary habits and their implications for platelet function. *Bull. Fac. Phys. Ther.* **2024**, 29, 7. [CrossRef]

- 2. Fabricius, H.-Å.; Starzonek, S.; Lange, T. The role of platelet cell surface P-selectin for the direct platelet-tumor cell contact during metastasis formation in human tumors. *Front. Oncol.* **2021**, *11*, 642761. [CrossRef]
- 3. Ho-Tin-Noe, B.; Boulaftali, Y.; Camerer, E. Platelets and vascular integrity: How platelets prevent bleeding in inflammation. *Blood* **2018**, *131*, 277–288. [CrossRef]
- 4. Davi, G.; Patrono, C. Platelet activation and atherothrombosis. N. Engl. J. Med. 2007, 357, 2482–2494. [CrossRef] [PubMed]
- Das, R.K.; Datta, T.; Biswas, D.; Duss, R.; O'Kennedy, N.; Duttaroy, A.K. Evaluation of the equivalence of different intakes of Fruitflow in affecting platelet aggregation and thrombin generation capacity in a randomized, double-blinded pilot study in male subjects. BMC Nutr. 2021, 7, 80. [CrossRef]
- 6. van der Meijden, P.E.; Heemskerk, J.W. Platelet biology and functions: New concepts and clinical perspectives. *Nat. Rev. Cardiol.* **2019**, *16*, 166–179. [CrossRef]
- 7. Qu, M.; Zou, X.; Fang, F.; Wang, S.; Xu, L.; Zeng, Q.; Fan, Z.; Chen, L.; Yue, W.; Xie, X.; et al. Platelet-derived microparticles enhance megakaryocyte differentiation and platelet generation via miR-1915-3p. *Nat. Commun.* **2020**, *11*, 4964. [CrossRef]
- 8. Menter, D.G.; Kopetz, S.; Hawk, E.; Sood, A.K.; Loree, J.M.; Gresele, P.; Honn, K.V. Platelet "first responders" in wound response, cancer, and metastasis. *Cancer Metastasis Rev.* **2017**, *36*, 199–213. [CrossRef]
- 9. Martin, J.F.; Kristensen, S.D.; Mathur, A.; Grove, E.L.; Choudry, F.A. The causal role of megakaryocyte–platelet hyperactivity in acute coronary syndromes. *Nat. Rev. Cardiol.* **2012**, *9*, 658–670. [CrossRef]
- 10. Johansson, P.; Ostrowski, S.; Secher, N. Management of major blood loss: An update. *Acta Anaesthesiol. Scand.* **2010**, *54*, 1039–1049. [CrossRef] [PubMed]
- 11. Gaertner, F.; Massberg, S. Blood coagulation in immunothrombosis-At the frontline of intravascular immunity. *Semin. Immunol.* **2016**, *28*, 561–569. [CrossRef]
- 12. Nurden, A.T. Platelets, inflammation and tissue regeneration. *Thromb. Haemost.* **2011**, 105, S13–S33. [CrossRef]
- 13. Xu, X.R.; Zhang, D.; Oswald, B.E.; Carrim, N.; Wang, X.; Hou, Y.; Zhang, Q.; Lavalle, C.; McKeown, T.; Marshall, A.H.; et al. Platelets are versatile cells: New discoveries in hemostasis, thrombosis, immune responses, tumor metastasis and beyond. *Crit. Rev. Clin. Lab. Sci.* 2016, 53, 409–430. [CrossRef]
- 14. Das, D.; Adhikary, S.; Das, R.K.; Banerjee, A.; Radhakrishnan, A.K.; Paul, S.; Pathak, S.; Duttaroy, A.K. Bioactive food components and their inhibitory actions in multiple platelet pathways. *J. Food Biochem.* **2022**, *46*, e14476. [CrossRef] [PubMed]
- 15. O'Kennedy, N.; Duss, R.; Duttaroy, A.K. Dietary Antiplatelets: A New Perspective on the Health Benefits of the Water-Soluble Tomato Concentrate Fruitflow<sup>®</sup>. *Nutrients* **2021**, *13*, 2184. [CrossRef]
- 16. Das, D.; Shruthi, N.R.; Banerjee, A.; Jothimani, G.; Duttaroy, A.K.; Pathak, S. Endothelial dysfunction, platelet hyperactivity, hypertension, and the metabolic syndrome: Molecular insights and combating strategies. *Front. Nutr.* **2023**, *10*, 1221438. [CrossRef]
- 17. Natarajan, A.; Zaman, A.G.; Marshall, S.M. Platelet hyperactivity in type 2 diabetes: Role of antiplatelet agents. *Diab. Vasc. Dis. Res.* **2008**, *5*, 138–144. [CrossRef] [PubMed]
- 18. Massberg, S.; Brand, K.; Gruner, S.; Page, S.; Muller, E.; Muller, I.; Bergmeier, W.; Richter, T.; Lorenz, M.; Konrad, I.; et al. A critical role of platelet adhesion in the initiation of atherosclerotic lesion formation. *J. Exp. Med.* 2002, 196, 887–896. [CrossRef] [PubMed]

Nutrients **2025**, 17, 3418 15 of 17

19. Kaplan, Z.S.; Jackson, S.P. The role of platelets in atherothrombosis. *Hematol. Am. Soc. Hematol. Educ. Program* **2011**, 2011, 51–61. [CrossRef]

- 20. Xu, X.R.; Yousef, G.M.; Ni, H. Cancer and platelet crosstalk: Opportunities and challenges for aspirin and other antiplatelet agents. *Blood* **2018**, *131*, 1777–1789. [CrossRef]
- 21. Koupenova, M.; Corkrey, H.A.; Vitseva, O.; Manni, G.; Pang, C.J.; Clancy, L.; Yao, C.; Rade, J.; Levy, D.; Wang, J.P.; et al. The role of platelets in mediating a response to human influenza infection. *Nat. Commun.* **2019**, *10*, 1780. [CrossRef]
- 22. Puurunen, M.K.; Hwang, S.J.; Larson, M.G.; Vasan, R.S.; O'Donnell, C.J.; Tofler, G.; Johnson, A.D. ADP Platelet Hyperreactivity Predicts Cardiovascular Disease in the FHS (Framingham Heart Study). *J. Am. Heart Assoc.* **2018**, *7*, e008522. [CrossRef]
- 23. Tomczynska, M.; Salata, I.; Bijak, M.; Saluk-Bijak, J. The potential contribution and role of a blood platelets in autoimmune thyroid diseases. *J. Cell. Mol. Med.* **2018**, 22, 6386–6390. [CrossRef]
- 24. Garcia-Leon, M.J.; Liboni, C.; Mittelheisser, V.; Bochler, L.; Follain, G.; Mouriaux, C.; Busnelli, I.; Larnicol, A.; Colin, F.; Peralta, M.; et al. Platelets favor the outgrowth of established metastases. *Nat. Commun.* **2024**, *15*, 3297. [CrossRef]
- 25. El Haouari, M.; Rosado, J.A. Platelet function in hypertension. Blood Cells Mol. Dis. 2009, 42, 38–43. [CrossRef] [PubMed]
- 26. Xu, C.; Song, Z.; Hu, L.T.; Tong, Y.H.; Hu, J.Y.; Shen, H. Abnormal platelet parameters in inflammatory bowel disease: A systematic review and meta-analysis. *BMC Gastroenterol.* **2024**, 24, 214. [CrossRef] [PubMed]
- 27. Harifi, G.; Sibilia, J. Pathogenic role of platelets in rheumatoid arthritis and systemic autoimmune diseases. Perspectives and therapeutic aspects. *Saudi Med. J.* **2016**, *37*, 354–360. [CrossRef]
- 28. Vannice, G.; Rasmussen, H. Position of the academy of nutrition and dietetics: Dietary fatty acids for healthy adults. *J. Acad. Nutr. Diet.* **2014**, 114, 136–153. [CrossRef]
- 29. Podgórska, K.; Derkacz, A.; Szahidewicz-Krupska, E.; Jasiczek, J.; Dobrowolski, P.; Radziwon-Balicka, A.; Skomro, R.; Szuba, A.; Mazur, G.; Doroszko, A. Effect of regular aerobic activity in young healthy athletes on profile of endothelial function and platelet activity. *Biomed. Res. Int.* 2017, 2017, 8715909. [CrossRef] [PubMed]
- 30. Di Francescomarino, S.; Sciartilli, A.; Di Valerio, V.; Di Baldassarre, A.; Gallina, S. The effect of physical exercise on endothelial function. *Sports Med.* **2009**, *39*, 797–812. [CrossRef]
- 31. Heber, S.; Volf, I. Effects of physical (in)activity on platelet function. Biomed. Res. Int. 2015, 2015, 165078. [CrossRef]
- 32. Dutta-Roy, A.K. Dietary components and human platelet activity. Platelets 2002, 13, 67–75. [CrossRef] [PubMed]
- 33. O'Kennedy, N.; Raederstorff, D.; Duttaroy, A.K. Fruitflow<sup>®</sup>: The first European Food Safety Authority-approved natural cardio-protective functional ingredient. *Eur. J. Nutr.* **2017**, *56*, 461–482.
- 34. Duttaroy, A.K.; Jorgensen, A. Effects of kiwi fruit consumption on platelet aggregation and plasma lipids in healthy human volunteers. *Platelets* **2004**, *15*, 287–292. [CrossRef] [PubMed]
- 35. Dutta-Roy, A.K.; Gordon, M.J.; Kelly, C.; Hunter, K.; Crosbie, L.; Knight-Carpentar, T.; Williams, B.C. Inhibitory effect of Ginkgo biloba extract on human platelet aggregation. *Platelets* **1999**, *10*, 298–305. [CrossRef] [PubMed]
- 36. Dutta-Roy, A.K.; Crosbie, L.; Gordon, M.J. Effects of tomato extract on human platelet aggregation in vitro. *Platelets* **2001**, 12, 218–227. [CrossRef]
- 37. Larson, M.K.; Shearer, G.C.; Ashmore, J.H.; Anderson-Daniels, J.M.; Graslie, E.L.; Tholen, J.T.; Vogelaar, J.L.; Korth, A.J.; Nareddy, V.; Sprehe, M.; et al. Omega-3 fatty acids modulate collagen signaling in human platelets. *Prostaglandins Leukot. Essent. Fat. Acids* **2011**, *84*, 93–98.
- 38. De Carvalho, C.C.; Caramujo, M.J. The various roles of fatty acids. Molecules 2018, 23, 2583. [CrossRef]
- 39. Sullivan, K.M. Open Source Statistics for Public Health. Available online: https://www.openepi.com/SampleSize/SSPropor.htm (accessed on 10 February 2022).
- 40. The Lithuanian National Olympic Committee (LTOK). Available online: https://www.ltok.lt/en (accessed on 12 December 2021).
- 41. Skernevičius, J.; Milašius, K.; Raslanas, A.; Dadelienė, R. Sporto treniruotė (Sports training). In *Sportininkų Gebėjimai ir jų Ugdymas* (*Skills and Training of Athletes*), 1st ed.; Čepulėnas, A., Saplinskas, J., Paulauskas, R., Eds.; Lithuanian University of Educational Sciences Press: Vilnius, Lithuania, 2011; pp. 165–217.
- 42. Deakin, V.; Kerr, D.; Boushey, C. Measuring nutritional status of athletes: Clinical and research perspectives. In *Clinical Sports Nutrition*, 5th ed.; Burke, L.M., Deakin, V., Eds.; McGraw-Hill: North Ryde, Australia, 2015; pp. 27–53.
- 43. Capling, L.; Beck, K.L.; Gifford, J.A.; Slater, G.; Flood, V.M.; O'Connor, H. Validity of dietary assessment in athletes: A systematic review. *Nutrients* 2017, 9, 1313. [CrossRef]
- 44. Barzda, A.; Bartkevičiūtė, R.; Viseckienė, V.; Abaravičius, A.J.; Stukas, R. Maisto Produktų ir Patiekalų Porcijų Nuotraukų Atlasas (Atlas of Foodstuffs and Dishes); Republican Nutrition Center, Vilnius University Faculty of Medicine: Vilnius, Lithuania, 2007; pp. 7–42. Available online: https://www.hi.lt/uploads/Products/product\_324/Maisto\_produkt\_ir\_patiekal\_porcij\_nuotrauk\_atlasas\_2007\_m..pdf (accessed on 28 April 2021).
- 45. Nutrition Surveys and Calculations. Available online: http://www.nutrisurvey.de/ (accessed on 21 January 2021).
- 46. Sučilienė, S.; Abaravičius, A. *Maisto Produktų Sudėtis (Food Product Composition)*; Ministry of Health of the Republic of Lithuania: Vilnius, Lithuania, 2002; pp. 10–315.

Nutrients **2025**, 17, 3418 16 of 17

47. Kerksick, C.M.; Arent, S.; Schoenfeld, B.J.; Stout, J.R.; Campbell, B.; Wilborn, C.D.; Taylor, L.; Kalman, D.; Smith-Ryan, A.E.; Kreider, R.B.; et al. International Society of Sports Nutrition position stand: Nutrient timing. *J. Int. Soc. Sports Nutr.* 2017, 14, 33. [CrossRef]

- 48. Aragon, A.A.; Schoenfeld, B.; Wildman, R.; Kleiner, S.; Vandusseldorp, T.; Taylor, L.; Earnest, C.P.; Arciero, P.J.; Wilborn, C.; Kalman, D.S.; et al. International Society of Sports Nutrition position stand: Diets and body composition. *J. Int. Soc. Sports Nutr.* **2017**, *14*, 16. [CrossRef]
- 49. Mountjoy, M.; Sundgot-Borgen, J.; Burke, L.; Ackerman, K.E.; Blauwet, C.; Constantini, N.; Lebrun, C.; Lundy, B.; Melin, A.; Meyer, N.; et al. International Olympic Committee (IOC) consensus statement on relative energy deficiency in sport (RED-S): 2018 update. *Int. J. Sport Nutr. Exerc. Metab.* 2018, 28, 316–331. [CrossRef]
- 50. Kerksick, C.M.; Wilborn, C.D.; Roberts, M.D.; Smith-Ryan, A.; Kleiner, S.M.; Jäger, R.; Collins, R.; Cooke, M.; Davis, J.N.; Galvan, E.; et al. ISSN exercise & sports nutrition review update: Research & recommendations. *J. Int. Soc. Sports Nutr.* **2018**, *15*, 8. [CrossRef] [PubMed]
- 51. Maughan, R.J.; Burke, L.M.; Dvorak, J.; Larson-Meyer, D.E.; Peeling, P.; Phillips, S.M.; Rawson, E.S.; Walsh, N.P.; Garthe, I.; Geyer, H.; et al. IOC consensus statement: Dietary supplements and the high-performance athlete. *Int. J. Sport Nutr. Exerc. Metab.* **2018**, 28, 104–125. [CrossRef] [PubMed]
- 52. Berger, M.E.; Smesny, S.; Kim, S.W.; Davey, C.G.; Rice, S.; Sarnyai, Z.; Schlögelhofer, M.; Schäfer, M.R.; Berk, M.; McGorry, P.D.; et al. Omega-6 to omega-3 polyunsaturated fatty acid ratio and subsequent mood disorders in young people with at-risk mental states: A 7-year longitudinal study. *Transl. Psychiatry* **2017**, 7, e1220. [CrossRef]
- 53. Harris, J.; Benedict, F. A Biometric Study of Basal Metabolism in Man; Lippincott: Philadelphia, PA, USA, 1919.
- 54. Thomas, D.T.; Erdman, K.A.; Burke, L.M. American college of sports medicine joint position statement. Nutrition and athletic performance. *Med. Sci. Sports Exerc.* **2016**, *48*, 543–568.
- 55. Ainsworth, B.E.; Haskell, W.L.; Herrmann, S.D.; Meckes, N.; Bassett, D.R.; Tudor-Locke, C.; Greer, J.L.; Vezina, J.; Whitt-Glover, M.C.; Leon, A.S. Compendium of physical activities: A second update of codes and MET values. *Med. Sci. Sports Exerc.* **2011**, 43, 1575–1581. [CrossRef]
- 56. FAO/WHO/UNU. *Human Energy Requirements: Principles and Definitions. Report of a Joint FAO/WHO/UNU Expert Consultation;* WHO: Geneva, Switzerland, 2001. Available online: https://www.fao.org/4/y5686e/y5686e00.htm (accessed on 19 December 2021).
- 57. EN ISO 13488:2000; Quality Systems—Medical Devices—Particular Requirements for the Application of EN ISO 9002 (Revision of EN 46002:1996) (Identical to ISO 13488:1996). ISO: Geneva, Switzerland, 2000.
- 58. Pratiwi, M.P.; Syakdiyah, N.H.; Sintoro, H.P. Platelet-to-lymphocyte ratio (PLR), platelet-to-hemoglobin ratio (PHR) and systemic immune-inflammation index (SII) as predictive factors of amputation among Diabetic Foot Ulcer (DFU) in Ibnu Sina Gresik Regional Public Hospital, East Java, Indonesia. *Indones. J. Biomed. Sci.* **2024**, *18*, 158–161. [CrossRef]
- 59. Abdul-Sahib, N.S.; Mahmood, M.M.; Ad'hiah, A.H. The significance of inflammatory markers derived from blood cell counts in predicting rheumatoid arthritis among the elderly. *Rev. Colomb. Reumatol.* **2024**, *RCREU-2157*, 1–11. [CrossRef]
- 60. Cohen, J. Statistical Power Analysis for the Behavioral Sciences; Routledge Academic: New York, NY, USA, 1988.
- 61. Carson, J.A.S.; Lichtenstein, A.H.; Anderson, C.A.; Appel, L.J.; Kris-Etherton, P.M.; Meyer, K.A.; Petersen, K.; Polonsky, T.; Van Horn, L.; American Heart Association Nutrition Committee of the Council on Lifestyle and Cardiometabolic Health; et al. Dietary cholesterol and cardiovascular risk: A science advisory from the American Heart Association. *Circulation* **2020**, *141*, e39–e53. [CrossRef] [PubMed]
- 62. Nunes, L.A.S.; Lazarim, F.L.; Brenzikofer, R.; Macedo, D.V. Applicability of the reference interval and reference change value of hematological and biochemical biomarkers to sport science. *Int. Perspect. Top. Sports Med. Sports Inj.* **2012**, *4*, 77–98.
- 63. Zareifar, S.; Farahmand Far, M.R.; Golfeshan, F.; Cohan, N. Changes in platelet count and mean platelet volume during infectious and inflammatory disease and their correlation with ESR and CRP. J. Clin. Lab. Anal. 2014, 28, 245–248. [CrossRef] [PubMed]
- 64. Garai, B.; Chatterjee, S.; Mondal, S.; Mondal, T. Effect of exercise on platelet variables: An overview. *Int. J. Phys. Educ. Sports Health* **2017**, *4*, 506–510.
- 65. Schumacher, Y.O.; Schmid, A.; Grathwohl, D.; Bültermann, D.I.R.K.; Berg, A. Hematological indices and iron status in athletes of various sports and performances. *Med. Sci. Sports Exerc.* **2002**, *34*, 869–875. [CrossRef]
- 66. Segal, J.B.; Moliterno, A.R. Platelet counts differ by sex, ethnicity, and age in the United States. *Ann. Epidemiol.* **2006**, *16*, 123–130. [CrossRef]
- 67. Al-Khelaifi, F.; Diboun, I.; Donati, F.; Botrè, F.; Alsayrafi, M.; Georgakopoulos, C.; Suhre, K.; Yousri, N.A.; Elrayess, M.A. A pilot study comparing the metabolic profiles of elite-level athletes from different sporting disciplines. *Sports Med. Open* **2018**, *4*, 2. [CrossRef]
- 68. Wang, C.; Harris, W.S.; Chung, M.; Lichtenstein, A.H.; Balk, E.M.; Kupelnick, B.; Jordan, H.S.; Lau, J. n-3 Fatty acids from fish or fish-oil supplements, but not alpha-linolenic acid, benefit cardiovascular disease outcomes in primary- and secondary-prevention studies: A systematic review. *Am. J. Clin. Nutr.* **2006**, *84*, 5–17. [CrossRef]

Nutrients 2025, 17, 3418 17 of 17

69. Andrioli, G.; Carletto, A.; Guarini, P.; Galvani, S.; Biasi, D.; Bellavite, P.; Corrocher, R. Differential effects of dietary supplementation with fish oil or soy lecithin on human platelet adhesion. *Thromb. Haemost.* **1999**, *82*, 1522–1527. [CrossRef]

- 70. Fischer, S.; Weber, P.C. Thromboxane A3 (TXA3) is formed in human platelets after dietary eicosapentaenoic acid (C20:5 omega 3). *Biochem. Biophys. Res. Commun.* **1983**, *116*, 1091–1099. [CrossRef]
- 71. Golanski, J.; Szymanska, P.; Rozalski, M. Effects of omega-3 polyunsaturated fatty acids and their metabolites on haemostasis—Current perspectives in cardiovascular disease. *Int. J. Mol. Sci.* **2021**, *22*, 2394. [CrossRef] [PubMed]
- 72. Mallick, R.; Basak, S.; Das, R.K.; Banerjee, A.; Paul, S.; Pathak, S.; Duttaroy, A.K. Fatty Acids and their proteins in adipose tissue inflammation. *Cell Biochem. Biophys.* **2024**, *82*, 35–51. [CrossRef] [PubMed]
- 73. Jäger, R.; Heileson, J.L.; Abou Sawan, S.; Dickerson, B.L.; Leonard, M.; Kreider, R.B.; Kerksick, C.M.; Cornish, S.M.; Candow, D.G.; Cordingley, D.M.; et al. International society of sports nutrition position stand: Long-chain omega-3 polyunsaturated fatty acids. *J. Int. Soc. Sports Nutr.* 2025, 22, 2441775. [CrossRef] [PubMed]
- 74. Hashimoto, M.; Hossain, S.; Shido, O. Docosahexaenoic acid but not eicosapentaenoic acid withstands dietary cholesterol-induced decreases in platelet membrane fluidity. *Mol. Cell. Biochem.* **2006**, 293, 1–8. [CrossRef] [PubMed]
- 75. Svaneborg, N.; Kristensen, S.D.; Hansen, L.M.; Bullow, I.; Husted, S.E.; Schmidt, E.B. The acute and short-time effect of supplementation with the combination of n-3 fatty acids and acetylsalicylic acid on platelet function and plasma lipids. *Thromb. Res.* 2002, 105, 311–316. [CrossRef]
- 76. Eschen, O.; Christensen, J.H.; De Caterina, R.; Schmidt, E.B. Soluble adhesion molecules in healthy subjects: A dose-response study using n-3 fatty acids. *Nutr. Metab. Cardiovasc. Dis.* **2004**, *14*, 180–185. [CrossRef]
- 77. Vanschoonbeek, K.; Feijge, M.A.; Paquay, M.; Rosing, J.; Saris, W.; Kluft, C.; Giesen, P.L.; de Maat, M.P.; Heemskerk, J.W. Variable hypocoagulant effect of fish oil intake in humans: Modulation of fibrinogen level and thrombin generation. *Arter. Thromb. Vasc. Biol.* **2004**, *24*, 1734–1740. [CrossRef]
- 78. Din, J.N.; Harding, S.A.; Valerio, C.J.; Sarma, J.; Lyall, K.; Riemersma, R.A.; Newby, D.E.; Flapan, A.D. Dietary intervention with oil rich fish reduces platelet-monocyte aggregation in man. *Atherosclerosis* **2008**, 197, 290–296. [CrossRef]
- 79. Vericel, E.; Calzada, C.; Chapuy, P.; Lagarde, M. The influence of low intake of n-3 fatty acids on platelets in elderly people. *Atherosclerosis* **1999**, 147, 187–192. [CrossRef]
- 80. Croset, M.; Vericel, E.; Rigaud, M.; Hanss, M.; Courpron, P.; Dechavanne, M.; Lagarde, M. Functions and tocopherol content of blood platelets from elderly people after low intake of purified eicosapentaenoic acid. *Thromb. Res.* **1990**, *57*, 1–12. [CrossRef]
- 81. Driss, F.; Vericel, E.; Lagarde, M.; Dechavanne, M.; Darcet, P. Inhibition of platelet aggregation and thromboxane synthesis after intake of small amount of icosapentaenoic acid. *Thromb. Res.* **1984**, *36*, 389–396. [CrossRef]
- 82. Guillot, N.; Caillet, E.; Laville, M.; Calzada, C.; Lagarde, M.; Vericel, E. Increasing intakes of the long-chain omega-3 docosahexaenoic acid: Effects on platelet functions and redox status in healthy men. *FASEB J.* **2009**, 23, 2909–2916. [CrossRef]
- 83. Duttaroy, A.K. Functional foods in preventing human blood platelet hyperactivity-mediated diseases—An updated review. *Nutrients* **2024**, *16*, 3717. [CrossRef]
- 84. Santhakumar, A.B.; Bulmer, A.C.; Singh, I. A review of the mechanisms and effectiveness of dietary polyphenols in reducing oxidative stress and thrombotic risk. *J. Hum. Nutr. Diet.* **2014**, 27, 1–21. [CrossRef] [PubMed]
- 85. Saputri, F.C.; Azmi, N.U.; Puteri, M.U.; Damayanti; Novita, V.; Marisi, G.; Oktavira, E.; Sari, A.N.; Ronaningtyas, K.; Herawati, E. High-fat diet enhances platelet activation and is associated with proprotein convertase subtilisin kexin 9: An animal study. *Nutrients* 2023, 15, 4463. [CrossRef]
- 86. Nagy, B., Jr.; Jin, J.; Ashby, B.; Reilly, M.P.; Kunapuli, S.P. Contribution of the P2Y12 receptor-mediated pathway to platelet hyperreactivity in hypercholesterolemia. *J. Thromb. Haemost.* **2011**, *9*, 810–819. [CrossRef]
- 87. Mariscal-Arcas, M.; Carvajal, C.; Monteagudo, C.; Lahtinen, J.; Fernández de Alba, M.C.; Feriche, B.; Olea-Serrano, F. Nutritional analysis of diet at base camp of a seven thousand-metre mountain in the Himalayas. *Rev. Andal. Med. Deporte* **2010**, *3*, 127–132.
- 88. Stukas, R.; Pečiukonienė, M.; Kemerytė-Riaubienė, E.; Baškienė, V. Some aspects of fat metabolism in an athlete's body. *Sport Sci.* **2009**, 2, 44–49.
- 89. Fung, T.T.; Rimm, E.B.; Spiegelman, D.; Rifai, N.; Tofler, G.H.; Willett, W.C.; Hu, F.B. Association between dietary patterns and plasma biomarkers of obesity and cardiovascular disease risk. *Am. J. Clin. Nutr.* **2001**, *73*, 61–67. [CrossRef] [PubMed]
- 90. Kennedy, A.; Martinez, K.; Chuang, C.C.; LaPoint, K.; McIntosh, M. Saturated fatty acid-mediated inflammation and insulin resistance in adipose tissue: Mechanisms of action and implications. *J. Nutr.* **2009**, 139, 1–4. [CrossRef]

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.