Expression levels of FBXW7 and MDM2 E3 ubiquitin ligases and their c-Myc and p53 substrates in patients with dysplastic nevi or melanoma

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Abstract. E3 ubiquitin ligases are of interest as drug targets due to their involvement in the regulation of the functions and interactions of several proteins. Various E3 ligase complexes are considered oncogenes or tumor suppressors associated with the development of melanoma. These proteins regulate the functions of various signaling pathways and proteins, such as p53 and Notch. The aim of the present study was to determine the expression levels of F-box and WD repeat domain-containing 7 (FBXW7), c-Myc, MDM2 and p53 proteins in samples from patients with dysplastic nevi or melanoma, and to evaluate their association with clinicopathological parameters and prognosis of the disease. Paraffin blocks with postoperative material from 100 patients diagnosed with dysplastic moles or melanoma were used in the present study. Tissue microarrays and immunohistochemistry were used to examine FBXW7, c-Myc, MDM2 and p53 protein expression. The results revealed that there was significantly lower FBXW7 expression in advanced melanoma compared with dysplastic nevus, melanoma in situ and stage pT1 melanoma (P<0.001). Additionally, there was a statistically significant association between the expression levels of FBXW7 and the morphological type of the tumor (P<0.001). In addition, there was a strong positive association between FBXW7 expression and the changes in c-Myc expression (P<0.02), and a strong trend was observed between decreased FBXW7 expression and a higher risk of death in patients, with the major factor in patient mortality being the stages of melanoma. Additionally, p53 expression was associated with the depth of melanoma invasion and the morphological type of the tumor. In summary, FBXW7 expression exhibited the highest statistically significant prognostic value and associations with advanced melanoma. As the majority of FBXW7 substrates are oncoproteins, their degradation by FBXW7 may highlight these proteins as potential targets for the treatment of melanoma.

Introduction

E3 ligases of the ubiquitin proteasome system are involved in the regulation of protein functions, stability and degradation; some of the components of the E3 ligase complexes are considered oncogenes or tumor suppressors with regards to the development of melanoma (1,2). Previous studies have focused on identifying gene products whose expression is altered as a disease progresses, as these may be novel molecular targets for the treatment of melanoma or new prognostic markers to track the course of the disease (1,3).

F-box and WD repeat domain-containing 7 (FBXW7) is a well-studied F-box-containing protein (4-6). F-box proteins are subunits of SCF-type E3 ligases that recognize the substrate, bind to it and target it for ubiquitination and degradation (4). It has been demonstrated that FBXW7 can act as a tumor suppressor by negatively regulating the expression levels of several protein oncogenes, including c-Myc, Notch, cyclin E and c-Jun (5-8). In melanoma, FBXW7 acts as a tumor suppressor. Studies have revealed that FBXW7 expression is significantly decreased in primary and metastatic melanoma samples compared with samples of dysplastic nevi, and that this decreased FBXW7 expression is associated with melanoma progression (5,9). However, the mechanism underlying the decrease in FBXW7 expression in tumors remains unclear.

One of the most important regulators of FBXW7 is p53 (10,11). p53 is a major tumor suppressor protein, with

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mutations detected in several types of human cancer, such as breast cancer, bone and soft tissue sarcoma, brain tumor, adrenocortical carcinoma, leukemia, stomach cancer and colorectal cancer (12-17). Mao *et al* (11) revealed that FBXW7 mediates the critical role of p53 in responding to DNA damage, suggesting that FBXW7 may be a p53-dependent gene tumor suppressor involved in tumor development. Further studies have demonstrated that FBXW7 expression may be restored by targeting the p53 signaling pathway (10,11).

c-Myc is a protein that is found in ~70% of all types of human cancer, including leukemia (18,19), sarcoma (20) and hepatocellular carcinoma (21,22). c-Myc is important for normal cell growth, and cellular Myc protein levels are tightly controlled (23,24). At least four different ubiquitin ligase complexes can target c-Myc for proteasomal degradation, including FBXW7 (23). Previous studies have revealed increased c-Myc expression in advanced and metastatic melanoma (25-29).

The E3 ubiquitin ligase MDM2 is a major negative regulator of the p53 tumor suppressor protein, and by suppressing p53, MDM2 promotes tumor development (30). In normal cells, the presence of MDM2 is essential for maintaining p53 protein expression at a basal level by regulating its ubiquitination and degradation in the 26S proteasome (31,32). In addition, MDM2 suppresses p53 function by directly binding to the transcriptional binding site of p53, thereby preventing its interaction with transcription factors (33-35). p53 and MDM2 interact and form a negative autoregulation loop in which elevated p53 transcriptional levels activate MDM2, which in turn decrease p53 levels (15,30,36).

The aim of the present study was to determine the protein expression levels of FBXW7, c-Myc, MDM2 and p53 in patients with dysplastic nevi or melanoma using tissue samples. Additionally, the associations between the expression levels of these proteins with clinicopathological parameters and prognosis of the disease were evaluated to determine whether these proteins may be used as prognostic factors for patients with melanoma, potentially allowing for improved modeling of effective personalized treatment of melanoma.

Materials and methods

Study population. The present study was performed on tissue microarray (TMA) sections obtained from paraffin blocks of postoperative material from 100 patients with dysplastic nevi or skin melanoma treated at the National Cancer Institute (Vilnius, Lithuania) between January 2013 and December 2018. Histological and immunohistochemical analysis was performed at the National Center of Pathology affiliated to Vilnius University Hospital SantarosKlinikos (Vilnius, Lithuania). The present study was approved by the Vilnius Regional Committee of Biomedical Research (approval no. 158200-16-878-387; approval date, 2016-12-13).

The present study included patients >18 years old with surgically removed and histologically confirmed dysplastic nevi or melanoma. The present study included 16 patients diagnosed with dysplastic nevi, 16 with *in situ* melanoma, 17 with pathological tumor (pT) 1 stage, 17 with pT2 stage, 17 with pT3 stage and 17 with pT4 stage melanoma. The following clinicopathological parameters of patients were evaluated: Sex, age, pT stage (37), morphological tumor type, ulceration and localization. Of the 100 cases under analysis, 39 were male and 61 were female, with a median age of 61 years (range, 21-92 years).

Immunohistochemical (IHC) analysis. The TMAs were constructed from 10% buffered formalin-fixed (at room temperature for ~ 24 h) paraffin-embedded tissue blocks; 2-mm diameter cores were punctured from the tumor block as randomly selected by a pathologist (1 core per patient), thus producing 2 TMAs constructed using the tissue arraying instrument (TMA Master; 3DHISTECH, Ltd.). The TMA blocks for immunohistochemistry were cut into $2-\mu$ m-thick sections and mounted on TOMO adhesion glass slides (Matsunami Glass Ind., Ltd.). The sections were deparaffinized and rehydrated in a descending alcohol series, and antigen retrieval for antibodies against p53 and c-Myc proteins was performed using DAKO PTLink system with EnVision FLEX Target buffer (pH 8.0) at 95°C for 20 min (both from Dako; Agilent Technologies, Inc.), while that for antibodies against FBXW7 and MDM2 proteins was performed using a Ventana Benchmark Ultra system with Cell Conditioning solution (pH 8.5) at 100°C for 36 min. (both from Ventana Medical Systems, Inc.; Roche Diagnostics). The sections were blocked with FLEX Peroxide Block (cat. no. SM801; Dako; Agilent Technologies, Inc.) at room temperature for 5 min for p53, 10 min for c-Myc and 8 min for FBXW7 and MDM2.Subsequently,the sections were incubated with antibodies against p53 (1:200; cat. no. M7001, clone DO-7; Dako; Agilent Technologies, Inc.) and c-Myc (1:40; cat. no. ab32072, cloneY69; Abcam) at room temperature for 30 min, then incubated using a DAKO EnVision FLEX system (Dako; Agilent Technologies, Inc.) for 20 min at room temperature. Incubation with antibodies against FBXW7 (1:50; cat. no. MA5-26562, clone OTI6F5; Thermo Fisher Scientific, Inc.) and MDM2 (1:250; cat. no. MA1-113, clone IF2; Invitrogen; Thermo Fisher Scientific, Inc.) was performed at 37°C for 32 min and then using the VentanaUltraview DAB detection kit (Ventana Medical Systems, Inc.; Roche Diagnostics) for 8 min at 37°C. Finally, the sections were developed using DAB, counterstained using Mayer's hematoxylin at room temperature for 10 min and mounted. Negative controls were performed by omitting the application of the primary antibody. The IHC slides were observed using a light microscope at a magnification of x20 (0.5 μ m resolution) using a bright field AperioScanScope XT Slide Scanner (Leica Microsystems, Inc.).

Assessment of FBXW7, c-Myc, MDM2 and p53 expression. The pathologist performed a visual assessment of staining intensity and overall percentage of cells staining. The intensity of protein immunostaining was scored as 0-3: 0, Negative staining; 1, weak staining; 2, moderate staining; and 3, strong staining. The percentage of nuclear staining was graded in 4 categories: 1, 0-25%; 2, 26-50%; 3, 51-75%; and 4, 76-100%. The combined score obtained by multiplying the staining intensity score with the staining percentage score was graded as follows: 0-6, Low expression; and 7-12, high expression.

Statistical analysis. Statistical analysis was performed using STATA v11.2 (StataCorp LP). Fisher's exact test was used to



Figure 1. Low FBXW7 expression is associated with melanoma progression. DN, dysplastic nevi; NI, non-invasive melanoma/melanoma *in situ*; FBXW7, F-box and WD repeat domain-containing 7; pT, pathological tumor stage.

evaluate the association between protein expression and the patient clinicopathological parameters. Univariate and multivariate Cox proportional and hazard regression analyses were performed to estimate the crude hazard ratios (HRs), adjusted HRs and 95% confidence intervals (CIs) of HRs. P<0.05 was considered to indicate a statistically significant difference.

Results

Patient characteristics. Based on the morphological type of melanoma, there were 48 cases of superficial melanoma, 27 cases of nodular melanoma and 9 cases of lentigomaligna. Of all the melanoma cases analyzed, 26 had melanoma depths of ≤ 1 mm and 42 cases had depths of invasion >1 mm, while 32 cases were non-invasive. Of the 68 melanoma cases ranging from pathological stages pT1-pT4, 22 were ulcerated. In 77% of the cases, the tumor was diagnosed in sun-exposed areas (Table I).

FBXW7 expression is associated with pathological stage, melanoma invasion depth and tumor morphological type. High FBXW7 expression was observed in 53% of samples, and low expression was observed in 47% of cases (Table I). There was a significant association between FBXW7 expression and pT stage. There was a significantly lower level of FBXW7 expression in advanced melanoma (pT3/pT4) compared with dysplastic nevi, melanoma in situ and pT1/pT2 stage melanoma (P<0.001; Fisher's exact test; Table I). Fig. 1 shows the proportion of high/low FBXW7 expression in dysplastic nevi, melanoma in situ, pT1/pT2 and pT3/pT4 melanoma. Fig. 2 shows the changes of immunostaining of FBXW7 protein depending on pT stage (pT1-pT4). There was strong FBXW7 immunostaining in pT1 melanoma, moderate staining in pT2, weak staining in pT3 and negative staining in pT4 melanoma (Fig. 2).

Additionally, there was a statistically significant association between FBXW7 expression and the morphological type of the tumor (P<0.001); high FBXW7 expression was observed in 93.7% of dysplastic nevus tissues, 77.8% of lentigomaligna cases and 54.2% of cases of superficial spreading melanoma, whereas 81.5% of nodular melanomas exhibited low FBXW7 expression (Table I). Furthermore, there was a statistically significant association between FBXW7 expression and the tumor invasion (P<0.001), with a depth $\leq 1 \text{ mm}$ observed in 76.9% of cases with high FBXW7 expression, whereas 90.5% of cases with melanoma invasion >1 mm exhibited low FBXW7 expression (Table I). There was no statistically significant association between FBXW7 expression and sex, age or tumor localization (Table I).

p53 expression is associated with the depth of melanoma invasion and the morphological type of the tumor. There was no statistically significant association between p53 expression and melanoma pT stage. Almost equal proportions of high and low expression levels of p53 were observed in dysplastic nevi and melanoma *in situ*. Higher proportions of high p53 expression were observed in stage pT1/pT2 and pT3/pT4 melanoma samples compared with high p53 expression in dysplastic nevi and melanoma *in situ* tissues (Fig. 3). High p53 expression was observed in 70-80% of melanoma samples ranging from stage pT1 to pT4, but p53 expression was not associated with pT stage (P=0.06; Table II). Fig. 4 shows the immunostaining of p53 protein in different pT melanoma stages (pT1-pT4). Additionally, there was no association between p53 expression with sex, age and tumor localization.

There was a statistically significant association between p53 expression and depth of melanoma invasion (P=0.02); when melanoma depth was ≤ 1 and >1 mm, high p53 expression was observed in 76.9 and 76.2% of cases, respectively, whereas a low p53 protein expression was detected in 53.1% of non-invasive tumors (Table II). There was also a statistically significant association between p53 expression and the morphological type of the tumor (P=0.01), with high p53 expression observed in 85.2% of nodular melanomas and in 68.8% of superficial spreading melanomas, while p53 expression in lentigomaligna tissues was mainly low (Table II).

c-Myc protein expression is not associated with advanced melanoma. Examination of c-Myc protein expression (Fig. 5) revealed that its expression was high in 36% of cases and low in 64% of cases. c-Myc protein expression was not significantly associated with advanced melanoma. Similarly, c-Myc expression was not associated with any clinicopathological parameters, including sex, age, morphological type of the tumor, depth of invasion and localization (Table III).

Table I. Association between FBXW7 protein expression and clinicopathological variables.

Table II. Association between p53 protein expression and clinicopathological variables.

Low

47 (47.0)

16 (41.0)

17 (27.9)

13 (29.5)

20 (35.7)

8 (50.0) 9 (56.3)

3 (17.7)

5 (29.4)

5 (29.4)

3 (17.7)

6 (23.1)

10 (23.8)

17 (53.1)

15 (31.2)

6 (66.7)

4 (14.8)

8 (50.0)

2 (15.4)

28 (36.4)

3 (30.0)

associated with advanced

P-value

0.20

0.52

0.06

 0.02^{a}

0.01^a

0.35

Cliniconsthelesion		FBXW7 e n (BXW7 expression, n (%)		Cliniconsthelogical		p53 expression, n (%)	
variables	Ν	High	Low	P-value	variables	Ν	High	Low
All cases	100	53 (53.0)	47 (47.0)	_	All cases	100	53 (53.0)	47 (47
Sex				0.41	Sex			
Male	39	23 (54.8)	16 (45.2)		Male	39	23 (59.0)	16 (41
Female	61	30 (49.2)	31 (50.8)		Female	61	44 (72.1)	17 (27
Age, years				0.69	Age, years			
≤58	44	25 (56.8)	19 (43.2)		≤58	44	31 (70.5)	13 (29
>58	56	28 (50.0)	28 (50.0)		>58	56	36 (64.3)	20 (35
pT stage				0.001ª	pT stage			
Dysplastic nevi	16	15 (93.7)	1 (6.25)		Dysplastic nevi	16	8 (50.0)	8 (50
pTis	16	14 (73.7)	2 (12.5)		pTis	16	7 (43.7)	9 (56
pT1	17	15 (88.2)	2 (11.8)		pT1	17	14 (82.3)	3 (17
pT2	17	6 (35.3)	11(64.7)		pT2	17	12 (70.6)	5 (29
pT3	17	1 (5.9)	16 (94.1)		pT3	17	12 (70.6)	5 (29
pT4	17	2 (11.8)	15 (88.2)		pT4	17	14 (82.3)	3 (17
Depth of invasion, mm				0.001ª	Depth of invasion, mm			
≤1	26	20 (76.9)	6 (23.1)		≤1	26	20 (76.9)	6 (23
>1	42	4 (9.5)	38 (90.5)		>1	42	32 (76.2)	10 (23
Non-invasive	32	29 (90.6)	3 (9.4)		Non-invasive	32	15 (46.9)	17 (53
Morphology				0.001ª	Morphology			
Superficial spreading	48	26 (54.2)	22 (45.8)		Superficial spreading	48	33 (68.8)	15 (31
Lentigomaligna	9	7 (77.8)	2 (22.2)		Lentigomaligna	9	3 (33.3)	6 (66
Nodular	27	5 (18.5)	22 (81.5)		Nodular	27	23 (85.2)	4 (14
Dysplastic nevi	16	15 (93.7)	1 (6.5)		Dysplastic nevi	16	8 (50.0)	8 (50
Site				0.54	Site			
Sun-protected	13	5 (38.5)	8 (61.5)		Sun-protected	13	11 (84.6)	2 (15
Sun-exposed	77	42 (54.5)	35 (45.5)		Sun-exposed	77	49 (63.6)	28 (36
Unknown	10	6 (60.0)	4 (4.0)		Unknown	10	7 (70.0)	3 (30
Ulceration in melanoma (pT1-pT4)	68			0.18	^a P<0.02. pT, pathological to	umor.		
Present	22	5 (22.7)	17 (77.3)					
Absent	46	19 (41.3)	27 (58.7)		MDM2 expression lev	els are	e not associ	iated wi

^aP<0.001. pT, pathological tumor; FBXW7, F-box and WD repeat domain-containing 7.

Decreased c-Myc expression was observed in 56% of melanoma cases, and in 71.4% of these cases, decreased expression was observed at melanoma depths >1 mm. There were no changes in c-Myc expression in dysplastic nevus, as both increased and decreased expression was observed in 50% of cases. There was a slight decrease in c-Myc expression observed in cases with melanoma in situ and stage pT1 melanoma, and low c-Myc protein expression levels were observed in 70.6% of melanoma cases in stages pT2, pT3 and pT4. However, there was no statistically significant association between c-Myc expression levels and the pathological stage of melanoma (P=0.75).

clinicopathological parameters (Table IV and Fig. 6). FBXW7 protein expression is associated with c-Myc protein expression. Fisher's exact test was performed to evaluate the association between FBXW7, c-Myc, MDM2 and p53 expression. The results revealed that there was a statistically significant association between FBXW7 and c-Myc expression, revealing that 76.6% of cases with low FBXW7 expression had also low c-Myc expression (P=0.02; Table V). There was no other statistically significant association between protein expression.

melanoma. The present study revealed that there was low

MDM2 expression in 97% of all investigated tumor samples; therefore, MDM2 expression did not exhibit any statistically significant association with advanced melanoma or any other

High FBXW7 expression is positively associated with survival. A univariate Cox regression analysis was used to determine



Figure 2. FBXW7 protein expression in cutaneous melanoma. Representative images of (A) strong FBXW7 immunostaining in pT1 melanoma, (B) moderate FBXW7 immunostaining in pT2 melanoma, (C) weak FBXW7 staining in pT3 melanoma and (D) negative FBXW7 staining in pT4 melanoma. Marked areas indicate FBXW7 staining in malignant melanocytes. Magnification, x100. FBXW7, F-box and WD repeat domain-containing 7; pT, pathological tumor stage.



Figure 3. p53 expression in DN, NI, pT1/pT2 and pT3/pT4 melanoma. DN, dysplastic nevi; NI, non-invasive melanoma/melanoma in situ; pT, pathological tumor stage.

which of the analyzed clinicopathological indicators may be associated with survival. The results revealed that sex, age, morphological type and the localization of tumors were not significantly associated with mortality. However, tumor ulceration was found to increase the risk of death by 2.79 times, although this rate was not statistically significant (P=0.06). Patient survival was significantly influenced by pT stage, where the risk of death increased by 5.6 times with advanced stage (P=0.03). Taking into account the impact of changes in the expression levels of FBXW7, c-Myc and p53 on the risk of death, the results revealed that high FBXW7 expression decreased the risk of death and improved survival (P=0.08), and high expression levels of p53 increased the risk of death by 2.48 times (P=0.10). However, these rates were not statistically significant. The results revealed that c-Myc expression was not significantly associated with mortality (Table VI). A multivariate Cox regression analysis revealed no statistically significant results (Table VI).



Figure 4. p53 protein expression in cutaneous melanoma. Representative images of (A) moderate p53 immunostaining in pT1 melanoma and (B) in pT2 melanoma, and (C) strong p53 immunostaining in pT3 and (D) in pT4 melanoma. Marked areas indicate p53 staining in malignant melanocytes. Magnification, x100. pT, pathological tumor stage.



Figure 5. c-Myc protein expression in cutaneous melanoma. Representative images of (A) high c-Mycimmunostaining in pT1 melanoma, (B) moderate c-Mycimmunostaining in pT2 melanoma and (C) in pT3 melanoma, and (D) weak c-Mycimmunostaining in pT4 melanoma. Marked areas indicate c-Myc staining in malignant melanocytes. Magnification, x100. pT, pathological tumor stage.

		c-Myc ex n (
variables	Ν	High	Low	P-value
All cases	100	36 (36.0)	64 (64.0)	-
Sex				0.29
Male	39	17 (43.6)	22 (56.4)	
Female	61	19 (31.1)	42 (68.9)	
Age, years				0.53
≤58	44	14 (31.8)	30 (68.2)	
>58	56	22 (39.3)	34 (60.7)	
oT stage				0.75
Dysplastic nevus	16	8 (50.0)	8 (50.0)	
pTis	16	6 (37.5)	10 (62.5)	
pT1	17	7 (41.2)	10 (58.8)	
pT2	17	5 (29.4)	12 (70.6)	
pT3	17	5 (29.4)	12 (70.6)	
pT4	17	5 (29.4)	12 (70.6)	
Depth of invasion, mm				0.37
≤1	26	10 (38.5)	16 (61.5)	
>1	42	12 (28.6)	30 (71.4)	
Non-invasive	32	14 (43.7)	18 (56.3)	
Morphology				0.57
Superficial spreading	48	17 (35.4)	31 (64.6)	
Lentigomaligna	9	3 (33.3)	6 (66.7)	
Nodular	27	8 (29.6)	19 (70.4)	
Dysplastic nevus	16	8 (50.0)	8 (50.0)	
Site				0.59
Sun-protected	13	5 (38.5)	8 (61.5)	
Sun-exposed	77	29 (37.7)	48 (62.3)	
Unknown	10	2 (20.0)	8 (80.0)	
oT, pathological tumor.				

Table III. Association between c-Myc protein expression and clinicopathological variables.

Table IV. Association between	MDM2 protein expression a	and
clinicopathological variables.		

Cliniconsthelegical		MDM2 n			
variables	Ν	High	Low	P-value	
All cases	100	3 (3.0)	97 (97.0)	-	
Sex				0.56	
Male	39	2 (5.1)	37 (94.9)		
Female	61	1 (1.1)	60 (98.9)		
Age, years				0.58	
≤58	44	2 (4.5)	42 (95.5)		
>58	56	1 (1.8)	55 (98.2)		
pT stage				0.99	
Dysplastic nevus	16	0 (0.0)	16 (100.0)		
pTis	16	1 (6.3)	15 (93.7)		
pT1	17	1 (5.9)	16 (94.1)		
pT2	17	0 (0.0)	17 (100.0)		
pT3	17	1 (5.9)	16 (94.1)		
pT4	17	0 (0.0)	17 (100.0)		
Depth of invasion, mm				0.99	
≤1	26	1 (3.8)	25 (96.2)		
>1	42	1 (2.4)	41 (97.6)		
Non-invasive	32	1 (3.1)	31 (96.9)		
Morphology				0.34	
Superficial spreading	48	1 (2.1)	47 (97.9)		
Lentigomaligna	9	1 (11.1)	8 (88.9)		
Nodular	27	1 (3.7)	26 (96.3)		
Dysplastic nevus	16	0 (0.0)	16 (100.0)		
Site				0.99	
Sun-protected	13	0 (0.0)	13 (100.0)		
Sun-exposed	77	3 (3.9)	74 (96.1)		
Unknown	10	0 (0.0)	10 (100.0)		
pT, pathological tumor.					

Discussion

FBXW7 is a tumor suppressor that controls the protein expression levels of several oncogenes, including c-Myc, Notch, Cyclin E, c-Jun, Mcl-1 and m-TOR (5,38,39). However, little is known regarding the regulation of FBXW7 in tumors. Regulation of FBXW7 expression may occur at the transcriptional or protein levels, as well as by post-translational modifications, such as phosphorylation (40). p53 molecules, NUMB4, NF- κ B1, microRNA-27 and microRNA-223 are known to be important in FBXW7 regulation (10).

Previous studies have revealed decreased FBXW7 activity in melanoma cells (3,9). The present study demonstrated that FBXW7 expression was lower in primary melanoma compared with dysplastic nevi. FBXW7 expression in metastatic melanoma was lower compared with in primary melanoma, and its decreased expression was associated with a less favorable 5-year survival rate (9). Furthermore, *in vitro* experiments have demonstrated that FBXW7 suppresses the migration of melanoma cells via the MAPK/ERK signaling pathway; therefore, suppression of FBXW7 in melanoma cells results in increased cell migration and stress fiber formation (9).

The results of the present study revealed that FBXW7 expression decreased in advanced melanoma, and a statistically significant association was found between the decrease in FBXW7 expression and the increasing pT stage of melanoma. Additionally, a trend was observed between decreased FBXW7 expression and an increased risk of death in patients, although this association was not significant.

Previous studies have demonstrated that decreased FBXW7 expression is associated with melanoma progression and the accumulation of c-Myc protein (3,9,41). c-Myc is one of the major targets of FBXW7, and c-Myc regulates the expression of >15% of the genes involved in processes of cell differentiation, proliferation, protein synthesis, metabolism and apoptosis; thus, impaired c-Myc function may underlie

pression levels	TI: 1								
-	High	Low	P-value	High	Low	P-value	High	Low	P-value
High	27 (75.0)	9 (25.0)	0.27						
Low	40 (62.5)	24 (37.5)							
High	2 (66.7)	1 (33.3)	0.99	2 (66.7)	1 (33.3)	0.29			
Low	34 (35.0)	36 (65.0)		34 (35.0)	36 (65.0)				
High	33 (62.3)	20 (37.7)	0.30	25 (47.2)	28 (52.8)	0.02ª	2 (3.8)	51 (96.2)	0.99
Low	34 (72.3)	13 (27.7)		11 (23.4)	36 (76.6)		1 (2.1)	46 (97.9)	
	High Low High Low High Low	High Low27 (75.0) 40 (62.5)High Low2 (66.7) 34 (35.0)High High Low33 (62.3) 34 (72.3)	High27 (75.0)9 (25.0)Low40 (62.5)24 (37.5)High2 (66.7)1 (33.3)Low34 (35.0)36 (65.0)High33 (62.3)20 (37.7)Low34 (72.3)13 (27.7)	High27 (75.0)9 (25.0)0.27Low40 (62.5)24 (37.5)High2 (66.7)1 (33.3)0.99Low34 (35.0)36 (65.0)High33 (62.3)20 (37.7)0.30Low34 (72.3)13 (27.7)	High Low27 (75.0) 40 (62.5)9 (25.0) 24 (37.5)0.27High Low2 (66.7) 34 (35.0)1 (33.3) 36 (65.0)0.99 34 (35.0)High High 33 (62.3)20 (37.7) 20 (37.7)0.30 0.3025 (47.2) 11 (23.4)	High 27 (75.0) 9 (25.0) 0.27 Low 40 (62.5) 24 (37.5) 40 High 2 (66.7) 1 (33.3) 0.99 2 (66.7) 1 (33.3) Low 34 (35.0) 36 (65.0) 34 (35.0) 36 (65.0) 34 (35.0) 36 (65.0) High 33 (62.3) 20 (37.7) 0.30 25 (47.2) 28 (52.8) Low 34 (72.3) 13 (27.7) 11 (23.4) 36 (76.6)	High Low $27 (75.0)$ $40 (62.5)$ $24 (37.5)0.27HighLow2 (66.7)4 (35.0)1 (33.3)36 (65.0)0.9934 (35.0)2 (66.7)34 (35.0)1 (33.3)36 (65.0)HighHighLow33 (62.3)20 (37.7)0.3011 (23.4)25 (47.2)36 (76.6)0.2911 (23.4)$	High Low $27 (75.0)$ $9 (25.0)$ 0.27 High $2 (66.7)$ $24 (37.5)$ $1 (33.3)$ 0.99 $2 (66.7)$ $1 (33.3)$ 0.29 Low $34 (35.0)$ $36 (65.0)$ $34 (35.0)$ $36 (65.0)$ $36 (65.0)$ High $33 (62.3)$ $20 (37.7)$ 0.30 $25 (47.2)$ $28 (52.8)$ 0.02^a $2 (3.8)$ Low $34 (72.3)$ $13 (27.7)$ $11 (23.4)$ $36 (76.6)$ $1 (2.1)$	High Low $27 (75.0)$ $9 (25.0)$ 0.27 Low $40 (62.5)$ $24 (37.5)$ High Low $2 (66.7)$ $1 (33.3)$ 0.99 $2 (66.7)$ $1 (33.3)$ 0.29 Low $34 (35.0)$ $36 (65.0)$ $34 (35.0)$ $36 (65.0)$ High High $33 (62.3)$ $20 (37.7)$ 0.30 $25 (47.2)$ $28 (52.8)$ 0.02^a $2 (3.8)$ $51 (96.2)$ Low $34 (72.3)$ $13 (27.7)$ $11 (23.4)$ $36 (76.6)$ $1 (2.1)$ $46 (97.9)$

Table V. Association between the changes in the protein expression levels of FBXW7, c-Myc, MDM2 and p53.

^aP<0.02. FBXW7, F-box and WD repeat domain-containing 7.



Figure 6. MDM2 protein expression in cutaneous melanoma. Representative images of (A) weak MDM2 immunostaining in pT1 melanoma and (B) in pT2 melanoma, (C) moderate MDM2 immunostaining in pT3 melanoma and (D) weak MDM2immunostaining in pT4 melanoma. Marked areas indicate MDM2 staining in malignant melanocytes. Magnification, x100. pT, pathological tumor stage.

tumor formation (5,42). FBW7 α promotes ubiquitination of Myc in proteasomes, whereas FBW7 γ ubiquitinates Myc in the nucleus and thus suppresses the ability of Myc to promote cell growth (41,43-46). Therefore, a decrease in FBXW7 expression results in increased c-Myc expression (10).

In the present study, 17 cases of stage pT2 melanoma, 17 cases of stage pT3 melanoma and 17 cases of stage pT4 melanoma were examined, and low c-Myc protein expression was detected in 12 cases (70.6%) in each group. According to a comparison of these groups with melanoma *in situ* and stage pT1 melanoma, the changes in c-Myc protein expression were not statistically significant. In addition, there was a strong direct association observed between the changes in FBXW7 and c-Myc expression with a decrease in FBXW7 expression, and a decrease in c-Myc expression was also observed. These results differ from those of previously published studies (41,45,47). The results of the present study may be influenced by the sample size. In addition, melanoma is a heterogeneous tumor and its development and progression is affected by the interaction of multiple genes and various signaling pathways (48). In some types of cancer, c-Myc may acquire loss-of-function mutations, while in the majority of cases, c-Myc expression is upregulated (49-52). In both cases, altered c-Myc expression results in tumor formation through disrupted transcription, translation or differentiation (42,49).

Variables	HR (95% CI)	P-value
Sex, males vs. females	2.01 (0.56-7.22)	0.29
Age, ≤58 vs. >58 years	1.16 (0.39-3.49)	0.79
pT stage, pT1/pT2 vs. pT3/pT4	5.60 (1.24-25.2)	0.03ª
Morphology, superficial	2.48 (0.83-7.43)	0.10
spreading vs. other		
Ulceration, absent vs. present	2.79 (0.95-8.08)	0.06
Site, sun-protected vs. sun-exposed	0.57 (0.16-2.11)	0.40
p53 expression, low vs. high	2.48 (0.83-7.43)	0.10
c-Myc expression, low vs. high	0.36 (0.08-1.63)	0.19
FBXW7 expression, low vs. high	0.16 (0.02-1.23)	0.08

tumor; FBXW7, F-box and WD repeat domain-containing 7.

Table VI. Univariate Cox regression analysis of protein expression and clinicopathological variables predicting the survival of patients with cutaneous melanoma.

Table VII. Multivariate Cox regression analysis of protein expression and clinicopathological variables predicting the survival of patients with cutaneous melanoma.

Variables	HR (95% CI)	P-value
Sex, males vs. females	1.47 (0.31-7.07)	0.48
Age, ≤58 vs. >58 years	0.69 (0.13-3.55)	0.66
pT stage, pT1/pT2 vs. pT3/pT4	3.13 (0.38-25.7)	0.29
Morphology, superficial	1.37 (0.28-6.60)	0.69
spreading vs. other		
Ulceration, absent vs. present	2.33 (0.60-9.09)	0.23
Site, sun-protected vs. sun-exposed	0.42 (0.09-1.98)	0.27
p53 expression, low vs. high	0.65 (0.16-2.70)	0.55
c-Myc expression, low vs. high	0.35 (0.07-1.72)	0.19
FBXW7 expression, low vs. high	0.65 (0.05-9.12)	0.75

^aP<0.05. HR, hazard ratio; CI, confidence interval; pT, pathological HR, hazard ratio; CI, confidence interval; pT, pathological tumor; FBXW7, F-box and WD repeat domain-containing 7.

One of the most important regulators of FBXW7 is p53; p53 is a major tumor suppressor protein and is frequently mutated in several types of cancer, such as breast cancer, bone and soft tissue sarcoma, brain tumor, adrenocortical carcinoma, leukemia, stomach cancer and colorectal cancer (12-17). Previous studies have demonstrated that FBXW7 is a p53-dependent tumor suppressor gene involved in tumor development (10,11). Additionally, studies have revealed that FBXW7 expression may be restored via targeting the p53 signaling pathway (10,11). The increased expression levels of wild-type p53 have been previously demonstrated in melanoma (53). However, considering the malignant nature and resistance to treatment in cases of melanoma (54,55), p53 does not appear to be effective as a tumor suppressor in melanoma. Although the mechanisms are not yet fully understood, certain p53 targets have been shown to be downregulated (56,57).

Increased expression levels of the MDM2 oncoprotein have been reported in several types of human cancer, including sarcoma, glioma, hematologic malignancies, melanoma and carcinoma, carrying the wild-type p53 allele (35,58). High levels of MDM2 are associated with a worse prognosis in several types of cancer, such as sarcoma, glioma and pediatric acute lymphoblastic leukemia (58,59). Malignant melanoma is characterized by increased MDM2 expression (60). Rajabi et al (61) revealed that there was an association between MDM2 expression with tumor thickness and invasion in primary cutaneous malignant melanoma. In 50% of melanoma cases, strong MDM2 expression is detected, leading to enhanced degradation of p53, and thus resulting in tumor cell proliferation (56,60,62).

In the present study, upregulated p53 expression was observed in primary and invasive melanoma. In cases of melanoma with a thickness >1 mm, p53 expression was increased compared with in non-invasive tumors, whereas FBXW7 expression decreased in advanced melanoma. Previous studies have demonstrated that the first exon of FBXW7 has a p53 binding site (11,63). Additionally, decreased FBXW7 expression following genotoxic stress may be activated by p53 (6,42). However, in the present study, there was no statistically significant association between p53 expression and the changes in FBXW7 expression.

p53 and MDM2 interact and form a negative autoregulatory loop in which elevated p53 transcriptional levels activate MDM2, which in turn decreases the levels of p53 (15,30,36). The present study revealed that there was a decrease in MDM2 expression in almost all cases assessed (97%). In contrast to published studies (60-62,64), the results of the present study did not identify a significant association between MDM2 expression and melanoma invasion, and there was no association between the decrease in MDM2 expression and p53 expression.

The main limitation of the present study consists in a relatively small number of subjects in the analyzed groups, assembled according to the stage of melanoma by depth. The sample size may have influenced the results obtained and the assessment of the effect of low FBXW7 protein expression on patient survival. Future studies should continue to analyze the expression levels of E3 ubiquitin ligases in melanoma tissues, since it is important to identify new potential targets for the treatment of melanoma, as well as to evaluate their prognostic significance. Future studies should investigate the expression levels of E3 ligases and their substrates at the mRNA level, as well as other genes involved in the development of melanoma, and should evaluate their interactions, their changes in expression associated with clinical characteristics and their prognostic significance.

In conclusion, the present study demonstrated that FBXW7 exhibited the most statistically significant prognostic value and associations with advanced melanoma. As most of the FBXW7 substrates are oncoproteins, their degradation by FBXW7 may underlie the mechanism by which decreased FBXW7 expression results in tumor progression and may highlight these proteins as potential targets for the treatment of melanoma.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

JM selected the patients, collected the clinicopathological data, interpreted the results and prepared the manuscript. ZG was involved in drafting and revising the manuscript, and made substantial contributions to the analysis and interpretation of data. IV performed the statistical analysis, interpreted the results and created the graphs and tables. AL was involved in performing the tissue microarray and immunohistochemical analysis, drafted and revised the manuscript. JP performed the histological examination of tissue samples of melanoma and dysplastic nevi, assisted in the tissue microarray preparation and evaluated the immunohistochemical staining. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The present study was approved by the Vilnius Regional Committee of Biomedical Research (approval no. 158200-16-878-387, approval date 2016-12-13, and addition no. 158200-878-PP1-05, 2017-03-17). All patients signed informed consent forms to participate in the present study (approval no. II-2016-5, 2016-12-07, version 4, and approval no. II-2016-5, 2017-02-14, version 5).

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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